

A toolkit of fluorescent molecular probes for biomembrane research

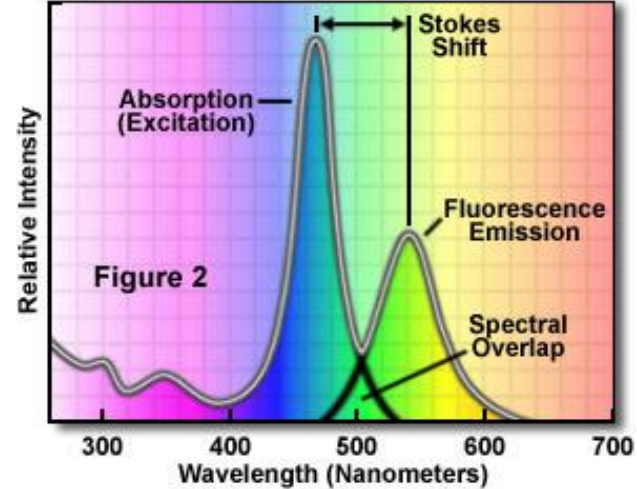
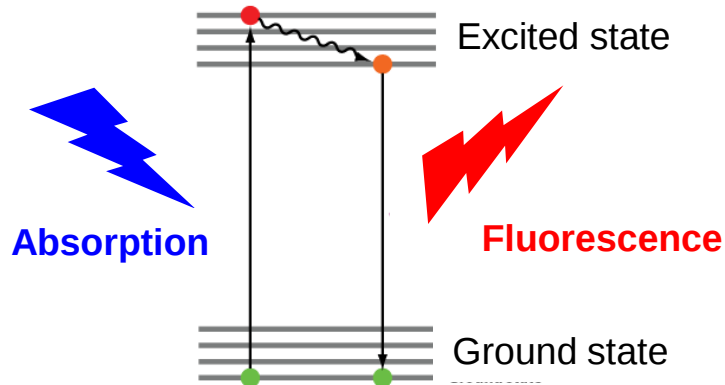
Andrey S. Klymchenko

Laboratoire de Biophotonique et Pharmacologie, UMR 7213 CNRS,
Université de Strasbourg, Faculté de Pharmacie, ILLKIRCH, France.

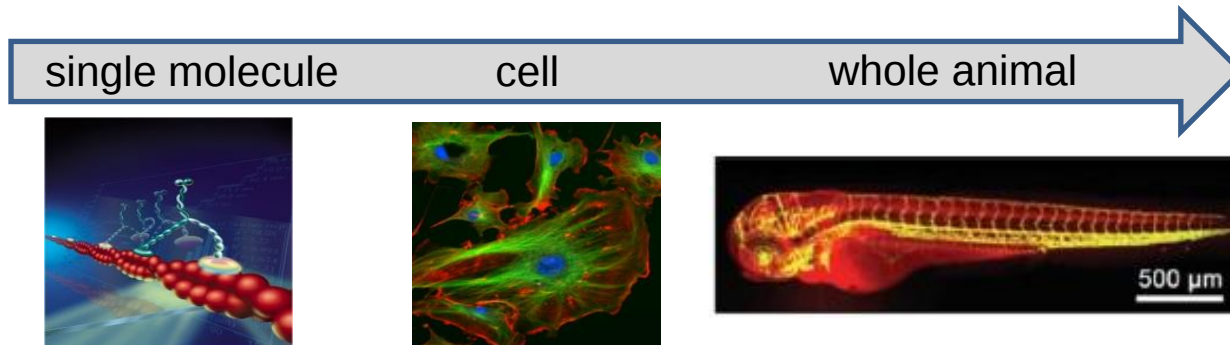
E-mail: andrey.klymchenko@unistra.fr



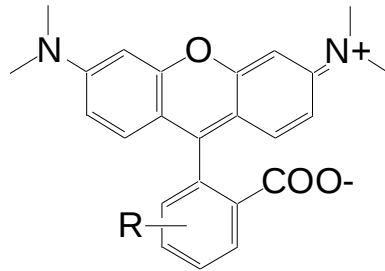
Why fluorescence?



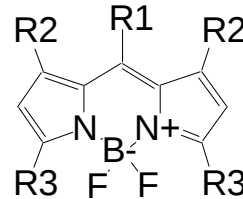
- **Sensitive:** up to single-molecule detection (10^{-6} - 10^{-12} M).
- **Fast:** any time scale is available up to picoseconds.
- **Non-invasive:** minimum effect on the object of study
- **Specific:** using specially designed molecule tools: fluorescent probes.



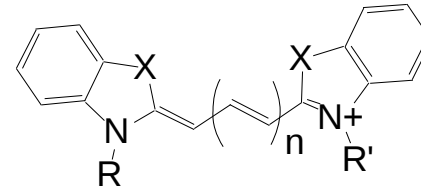
“Classical” fluorescent dyes



Rhodamine



BODIPY



Cyanine

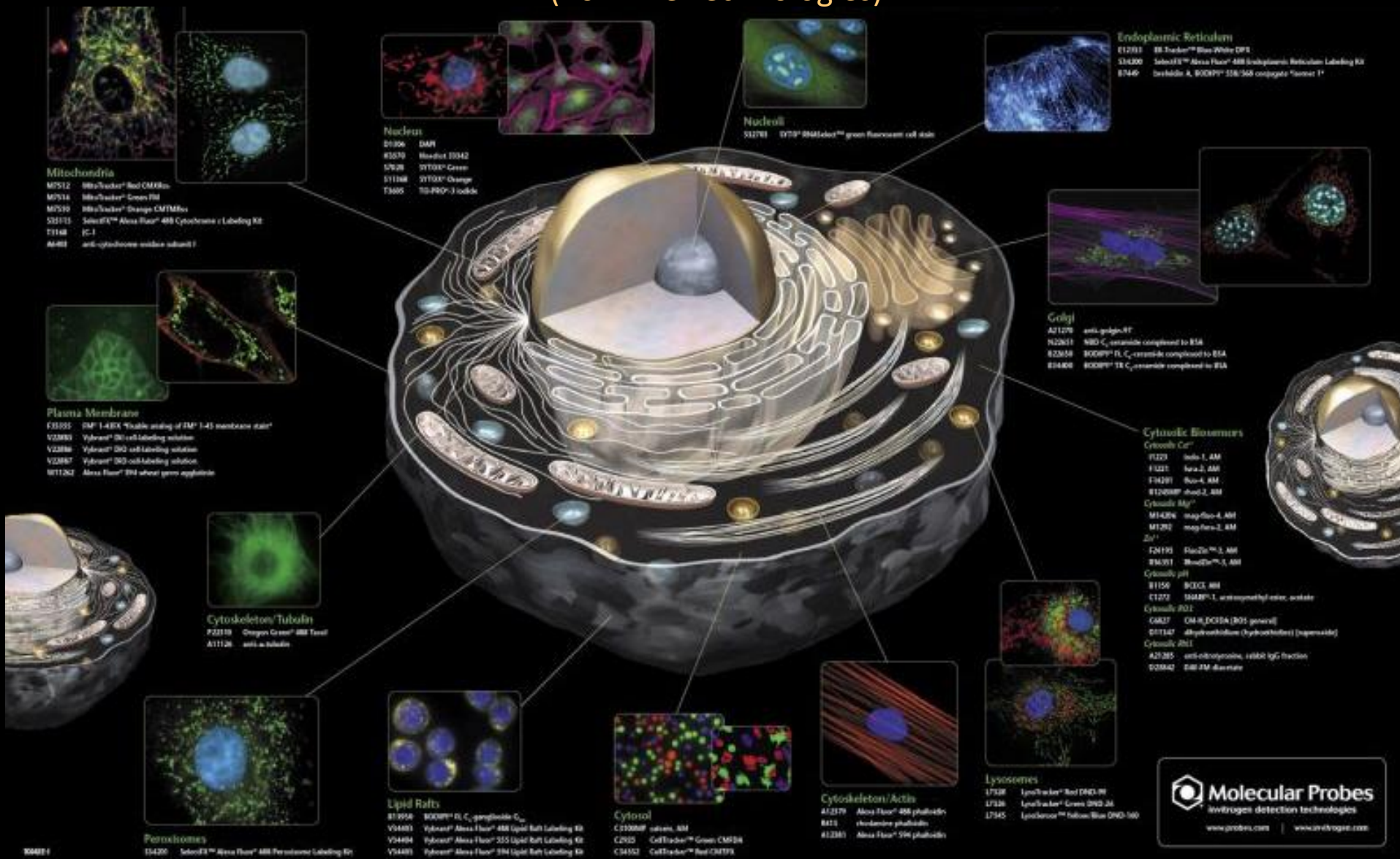
Cy3 (n=1)

Cy5 (n=2)

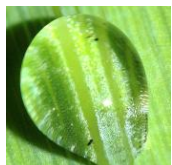
$\text{X} = \text{S}, \text{O}, \text{C}(\text{CH}_3)_2$

- Fluorescence of “classical” dyes is almost independent of the environment
- They are perfect markers, but they are not probes (unless sensor group is added).

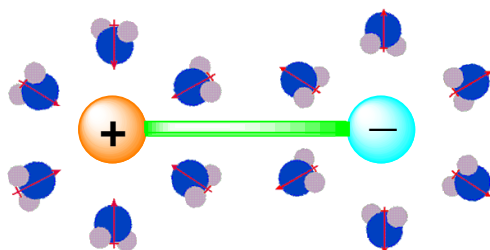
“The illuminated cell”: the successful project of Molecular Probes (now Life Technologies)



Environment-sensitive fluorescent dyes



Water:
Polar



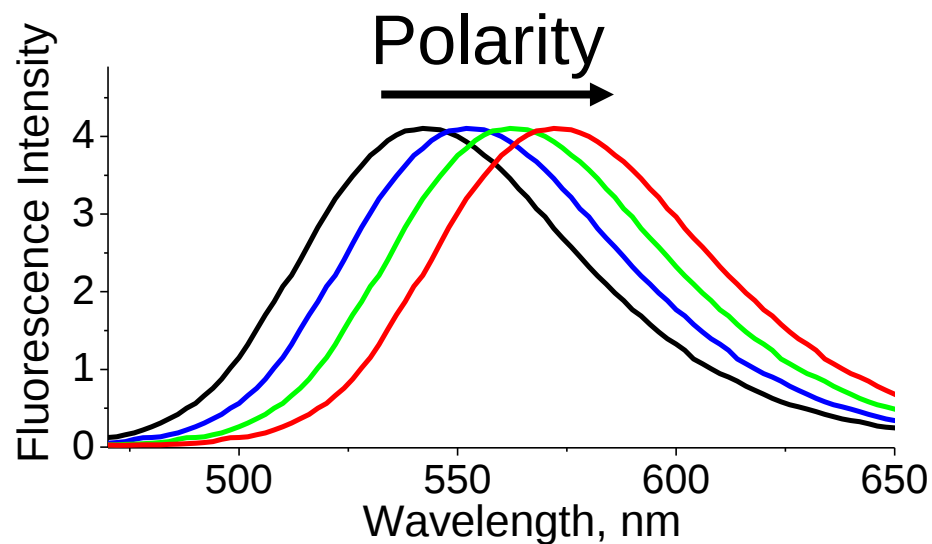
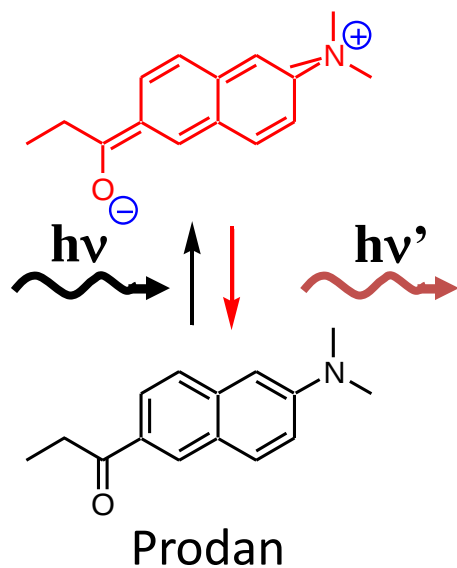
Strong solvation



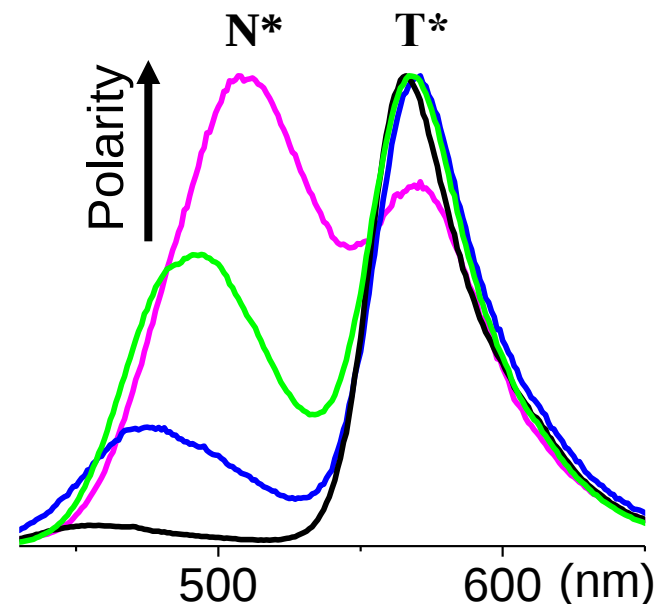
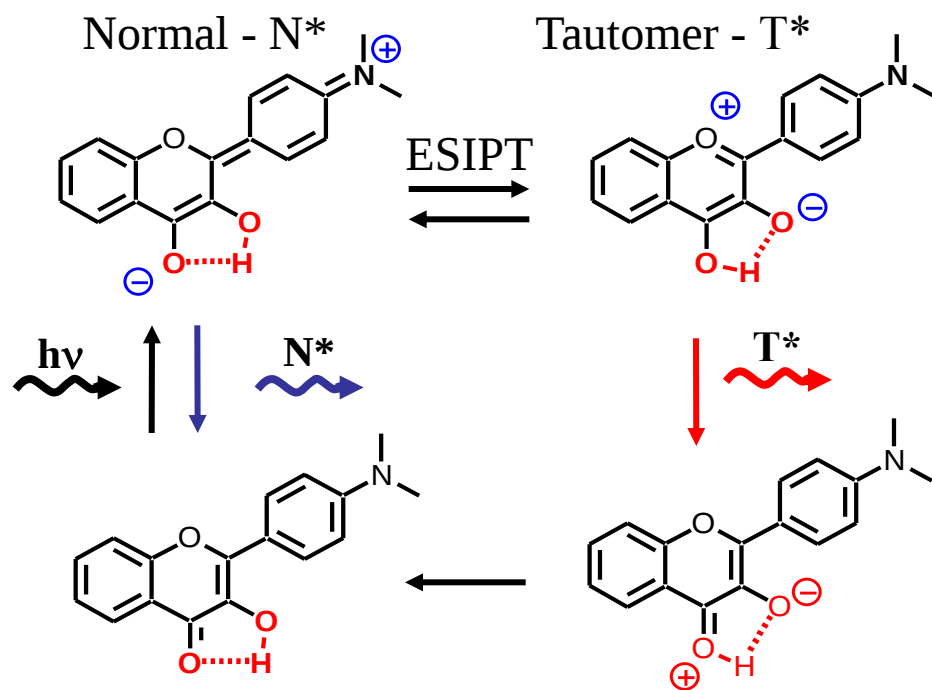
Oil:
nonpolar



Poor solvation



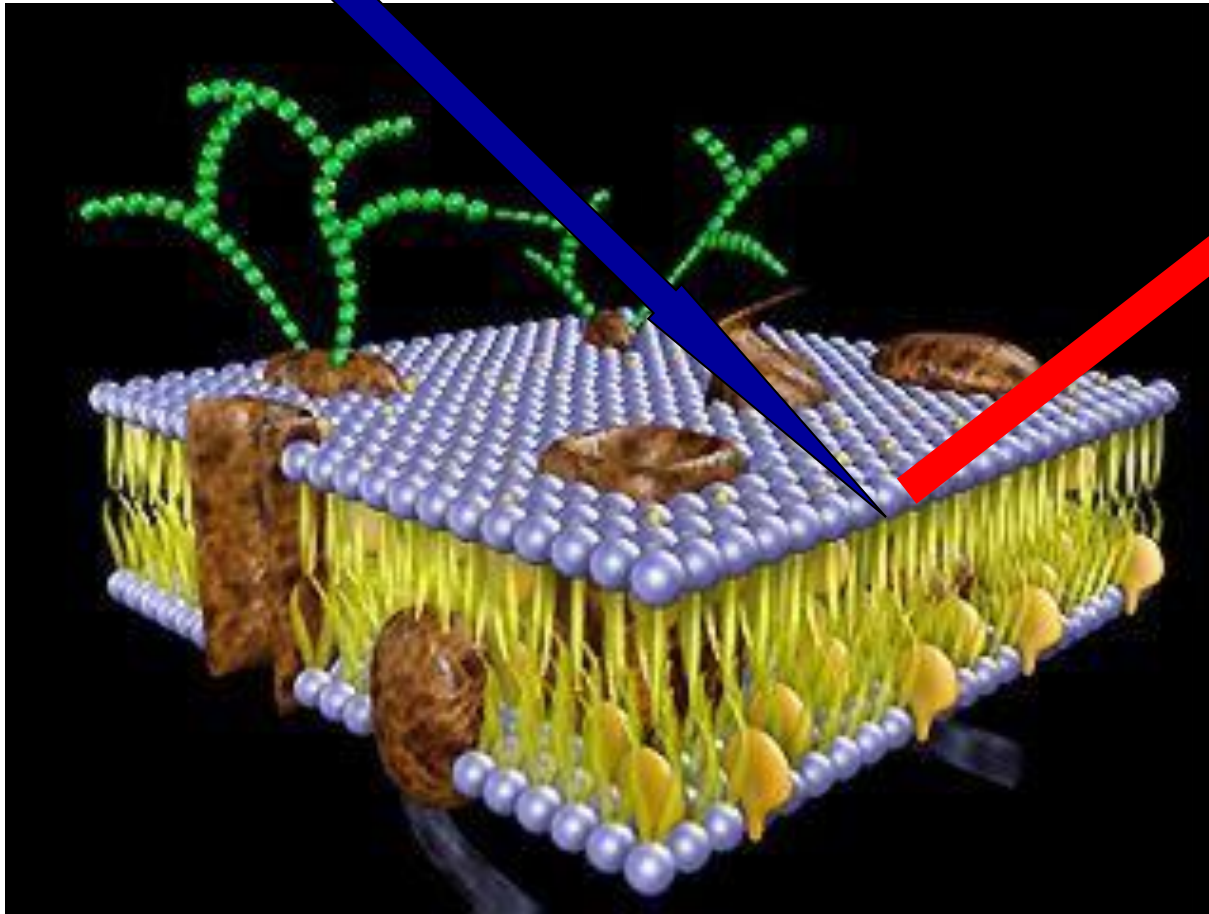
3-Hydroxychromones (3HCs): Solvatochromic ESIPT dyes



Advantages:

- 1) Satisfactory absorption properties: absorption ~ 400 nm ($35,000 \text{ M}^{-1}\text{cm}^{-1}$).
- 2) Fluorescence quantum yield: 5-50 %
- 3) Extreme sensitivity of dual emission to environment.
- 4) Additional channels of spectroscopic information.

Probes for membranes



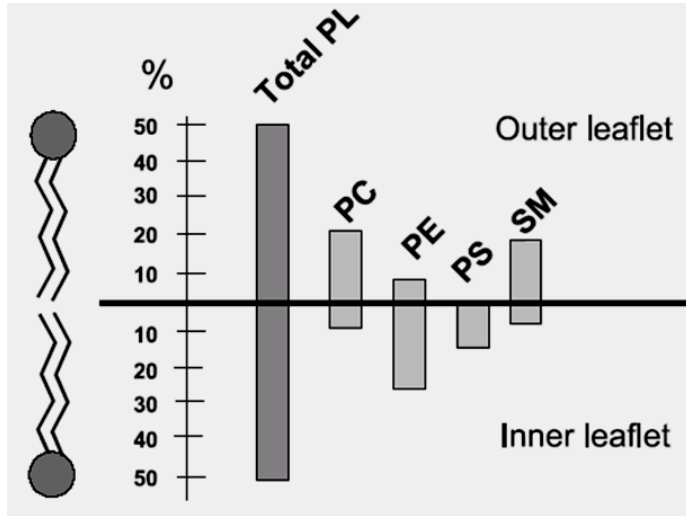
Fluorescence



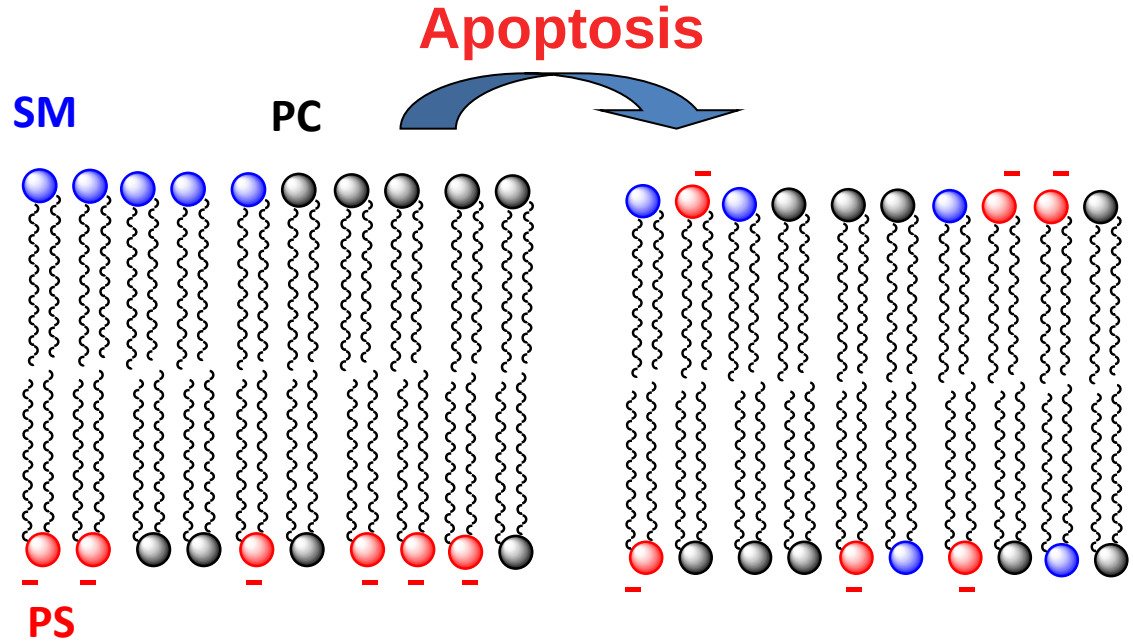
Membrane structure,
lipid distribution,
& phase state

Cellular membrane and apoptosis

Transmembrane asymmetry in plasma membranes



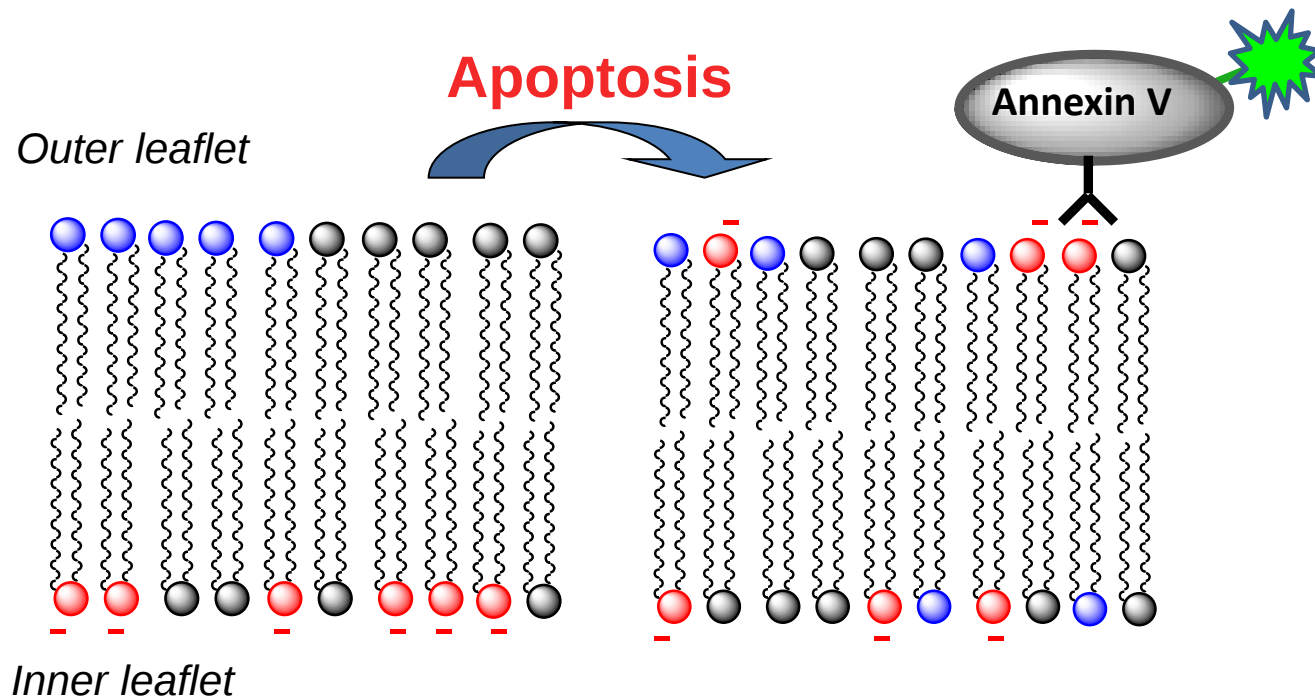
PC = phosphatidyl choline
PE = phosphatidyl ethanolamine
PS = phosphatidyl serine
SM = Sphingomyelin



The asymmetry is lost on apoptosis

- ✓ Surface charge increases
- ✓ Lipid order decreased?

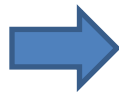
Cellular membrane and apoptosis



Van Engeland et al., *Cytometry* 1998.

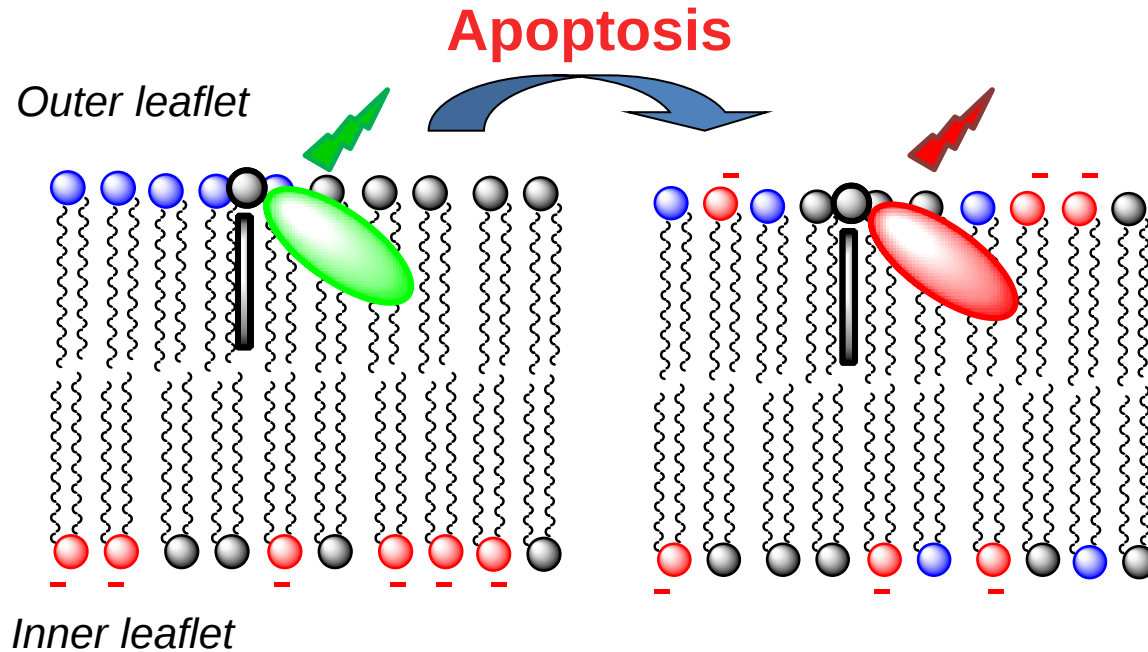
Some limitations of this assay:

- Ca^{2+} -dependent
- Expensive
- Intensiometric



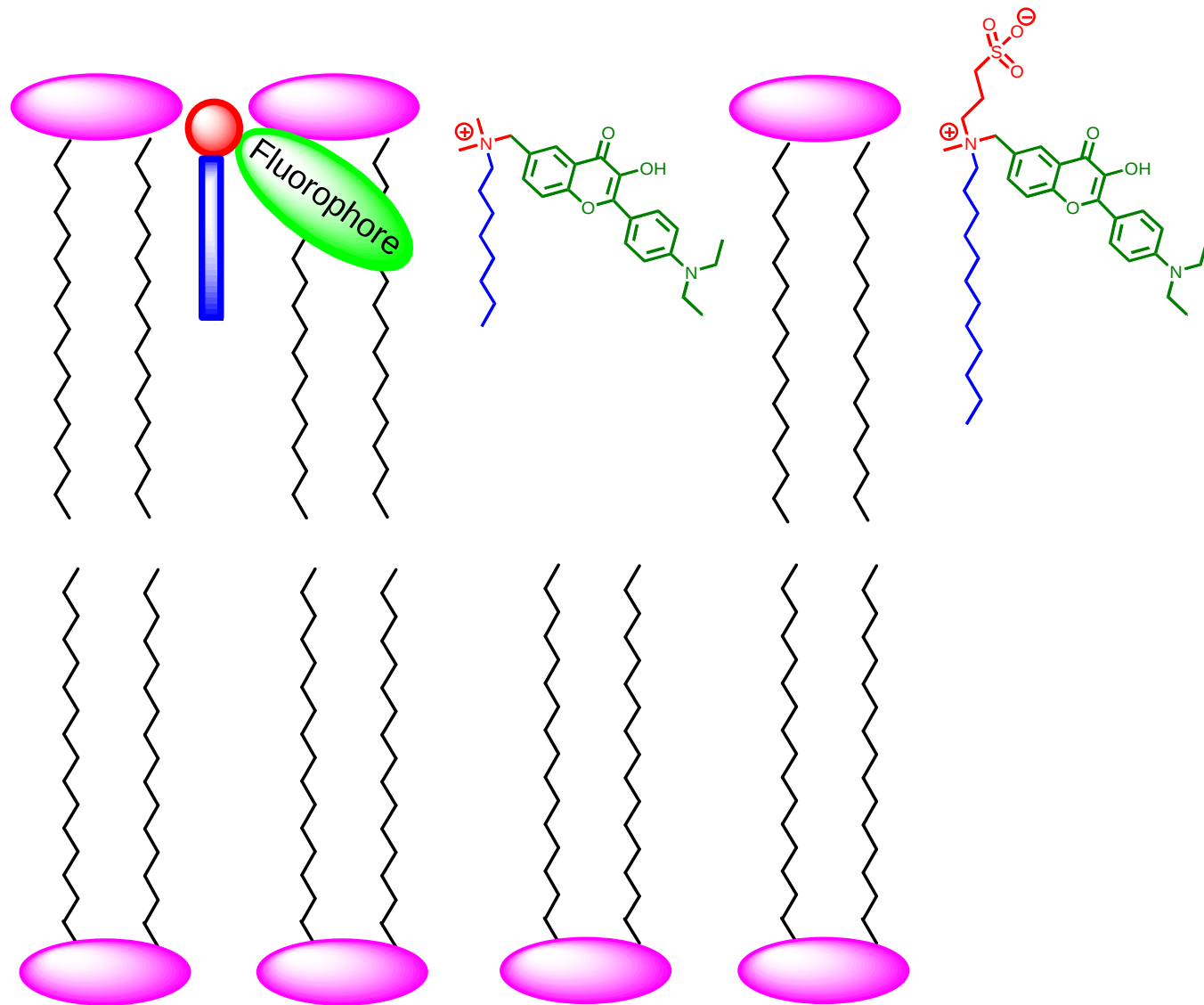
Alternative methods of apoptosis detection
based on membrane changes

Cellular membrane and apoptosis

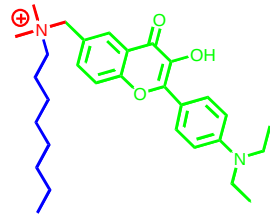


Dye should be sensitive to lipid composition:
surface charge and lipid order

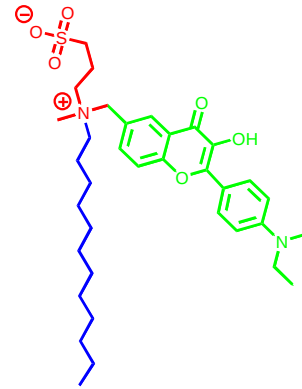
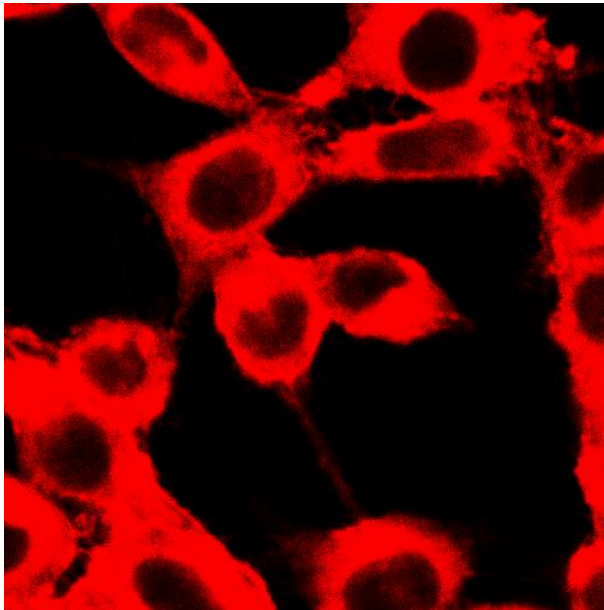
Localizing probe at the membrane



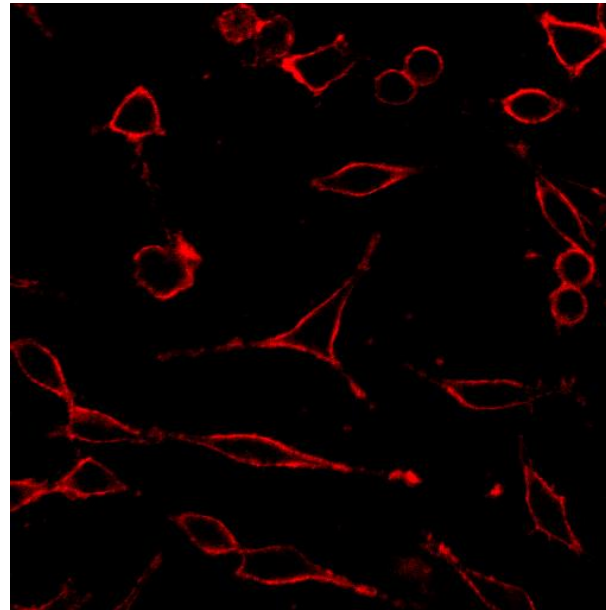
Design of the biomembrane probe



Internalize 😞



Stains only membrane 😊

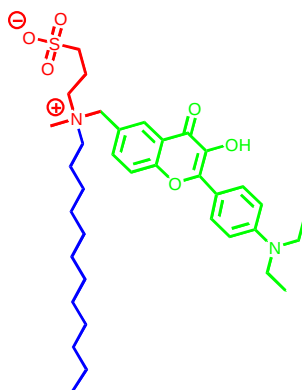
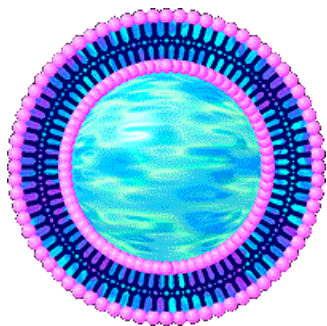


Confocal microscopy images

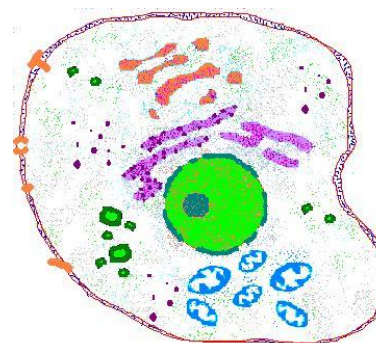
Fibroblast L 929 cell line stained with 1 μ M of probes

Response of F2N12S to surface charge in liposomes and to apoptosis in cells

In liposomes



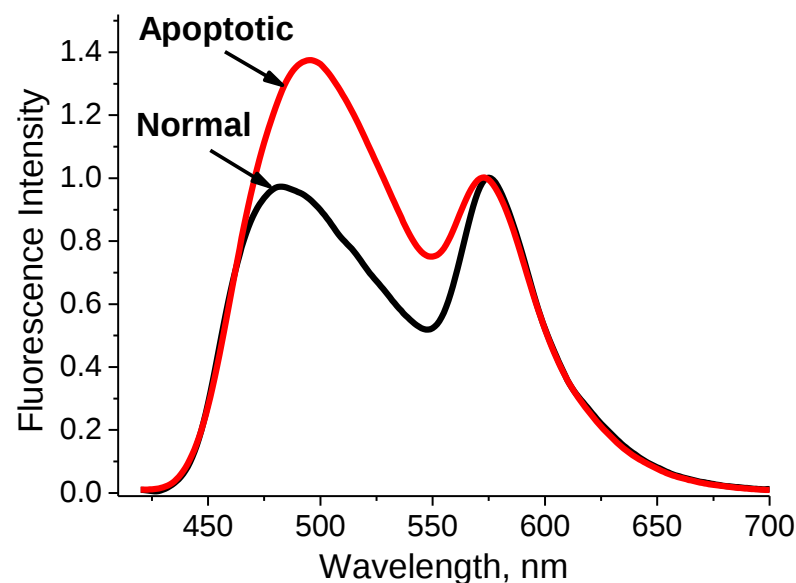
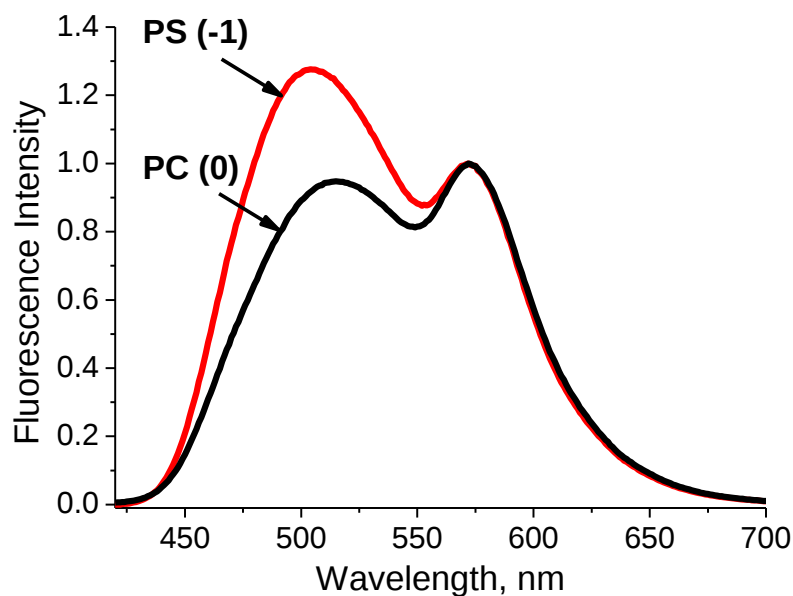
In cells



+actinomycin



Apoptosis

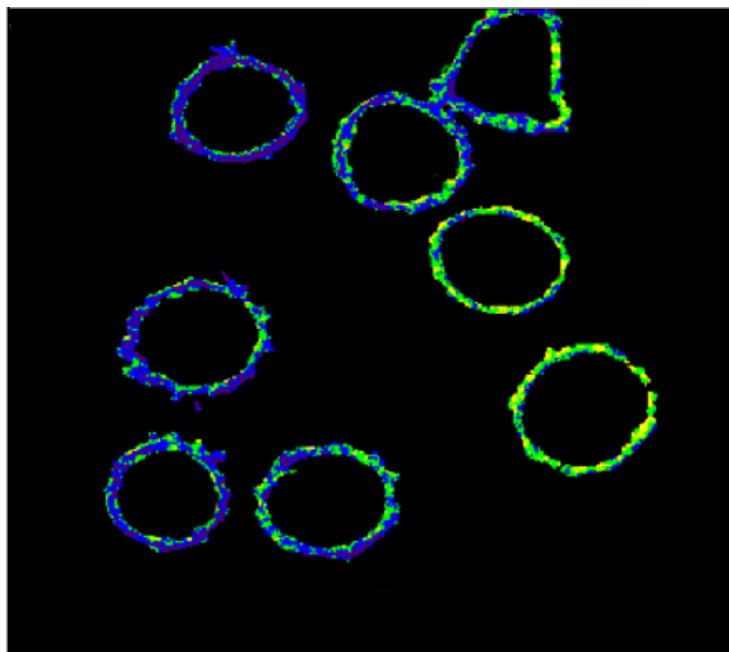


Fluorescence microscopy imaging of apoptosis with probe F2N12S

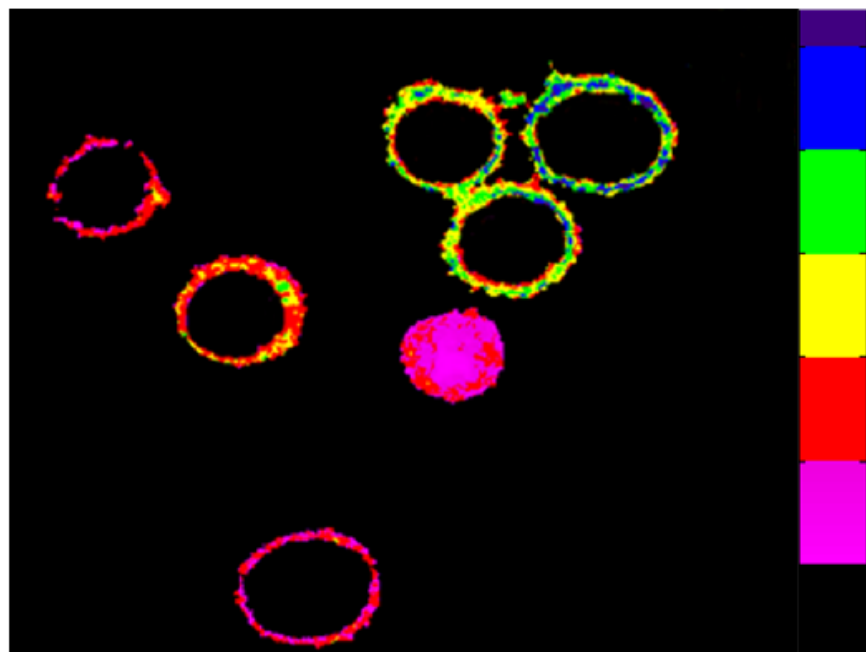
Apoptosis



$I_{\text{Red}} / I_{\text{Blue}}$



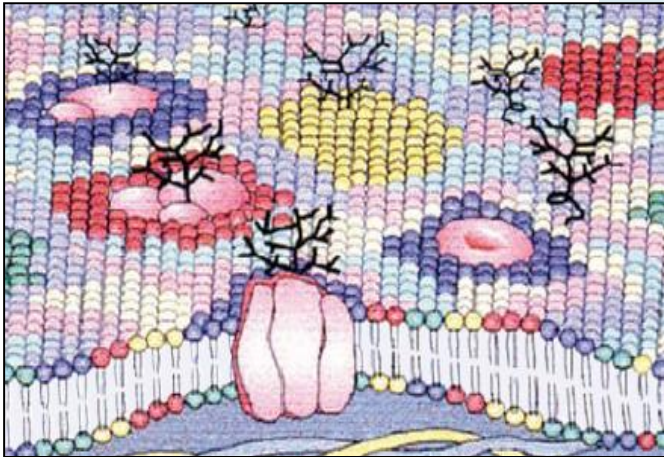
Non-treated



Treated with actinomycin for 18 h

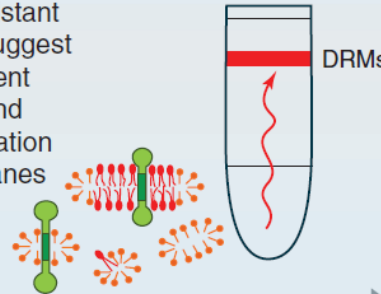
CEM cell line stained with 1 μM of probes

Hypothesis of "lipid rafts"



1990s

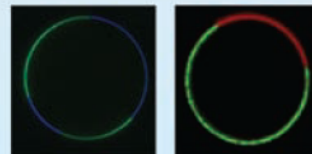
Detergent-resistant membranes suggest sterol-dependent sphingolipid and protein association in cell membranes



Antibody-patched proteins display selective sterol-dependent coalescence at the cell surface

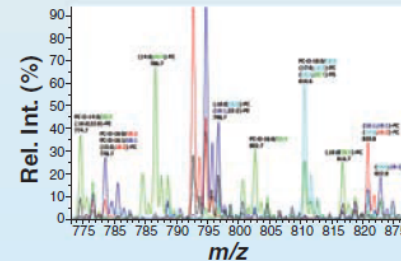
2000s

Macroscopic phases separate in model membranes and cell membranes



GUV

Plasma membrane



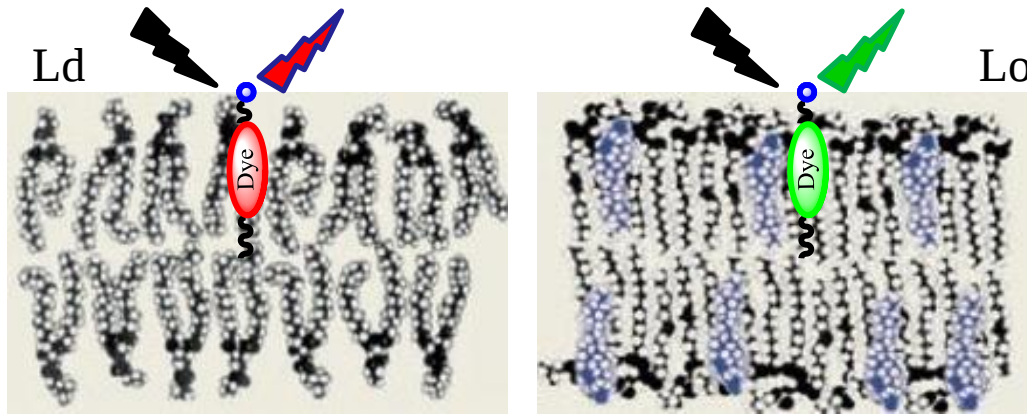
Lipidomics reveals that sphingolipids and sterol are sorted in the TGN during transport to the plasma membrane



Advances in microscopy and spectroscopy (e.g. SPT, FCS, FRET, STED, FPALM) reveal dynamic nanoassemblies of sterol, sphingolipid, and protein in living cells

Fluorescent probes for lipid rafts: what is needed?

Phase-sensitivity



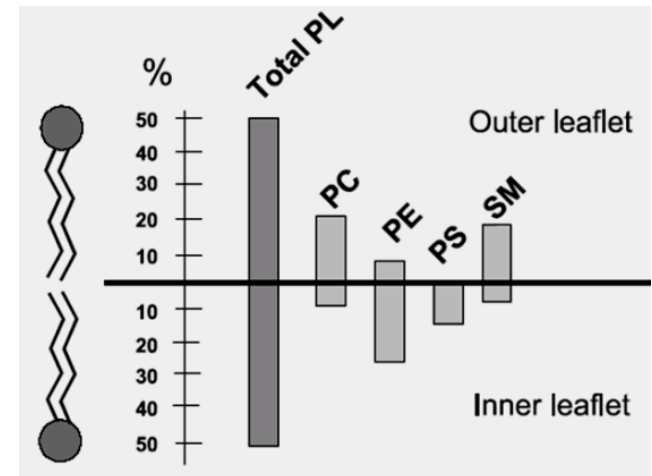
- ✓ Loosely packed
- ✓ Fast dynamics
- ✓ Hydrated
- ✓ Polar

- ✓ Tightly packed
- ✓ Slow dynamics
- ✓ Dehydrated
- ✓ Apolar



Use of environment-sensitive
(solvatochromic) probes?

Leaflet-specificity



- ✓ Order in outer & inner
leaflets is different

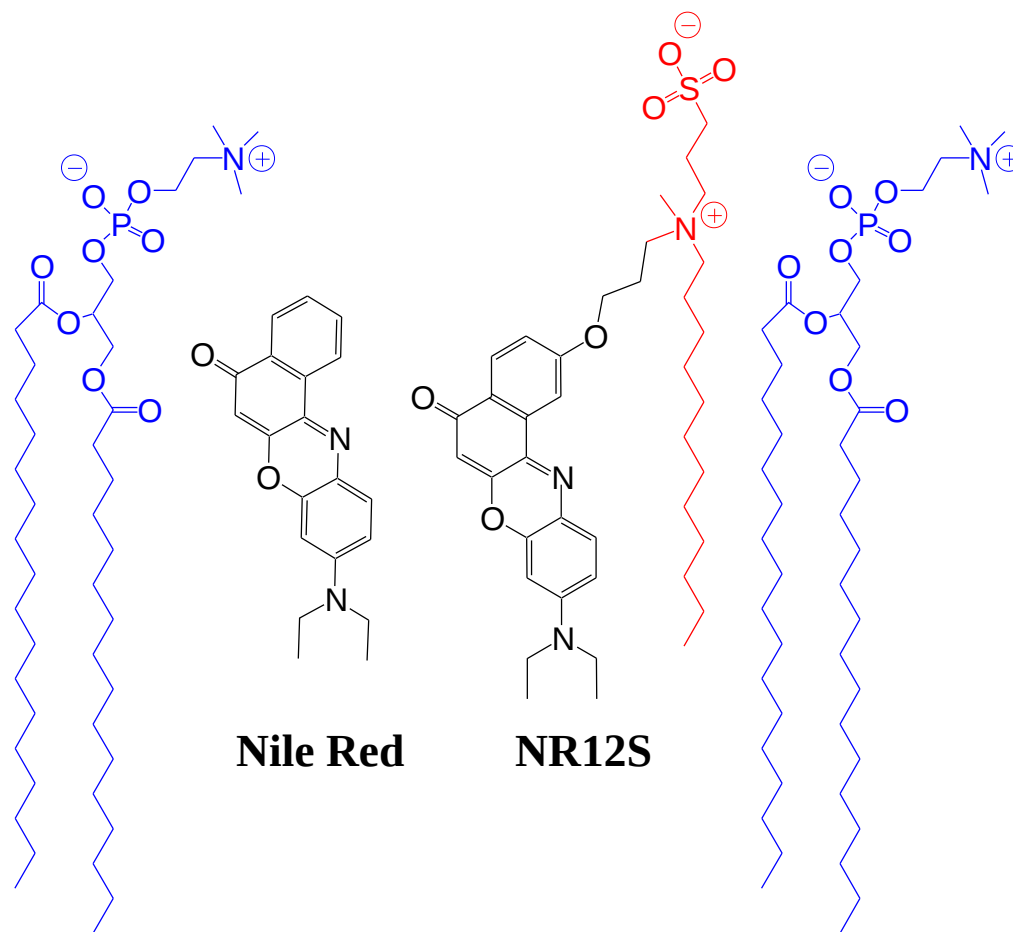
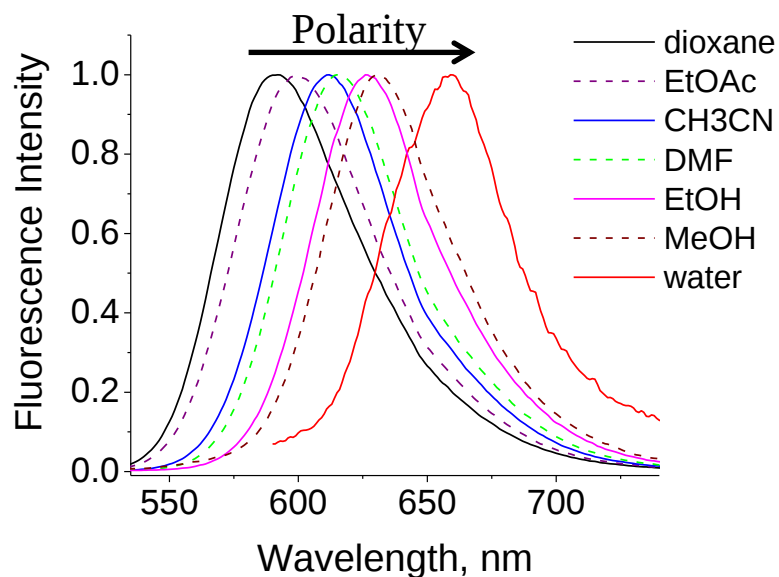


Specific staining of one leaflet

Nile Red-based probe

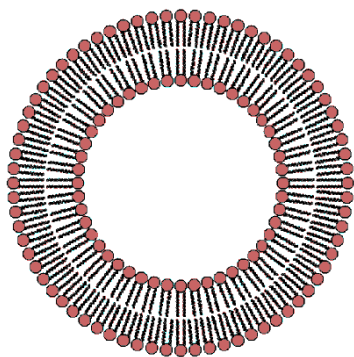
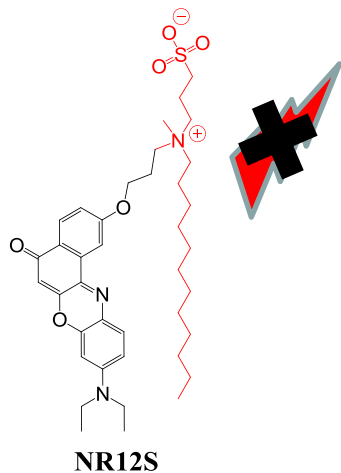
Key features of Nile Red:

- 1) Bright: $\epsilon = 40,000$; QY ~ 0.5 .
- 2) Absorption max at 550 nm.
- 3) Environment-sensitive

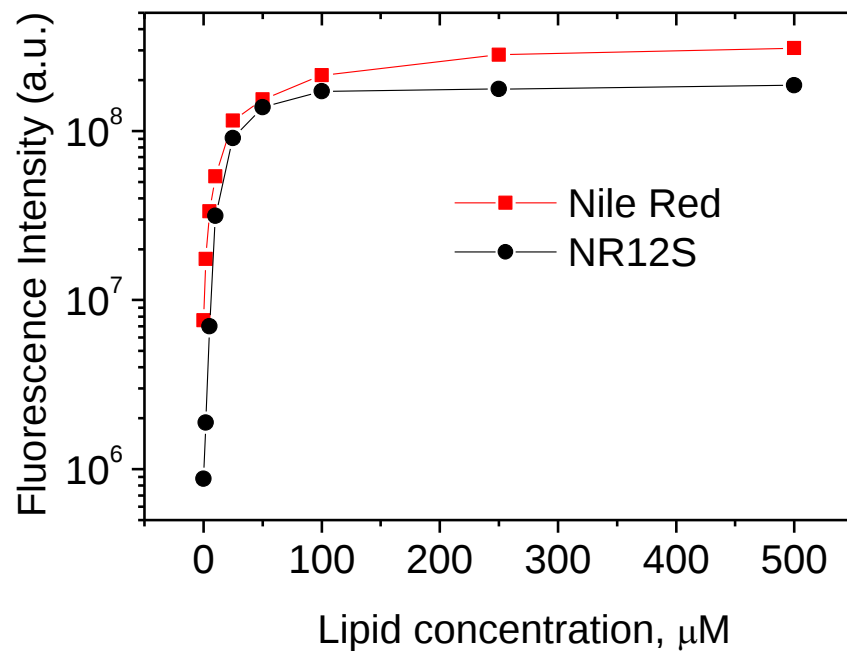
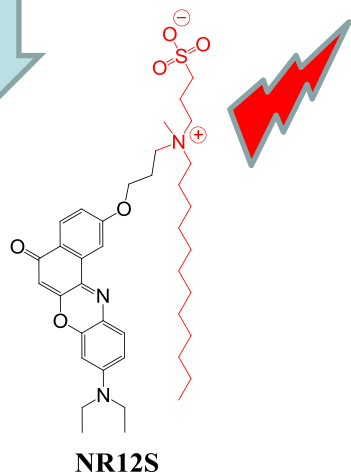


Fluorescence of NR12S "turns on" after binding to membranes

In water



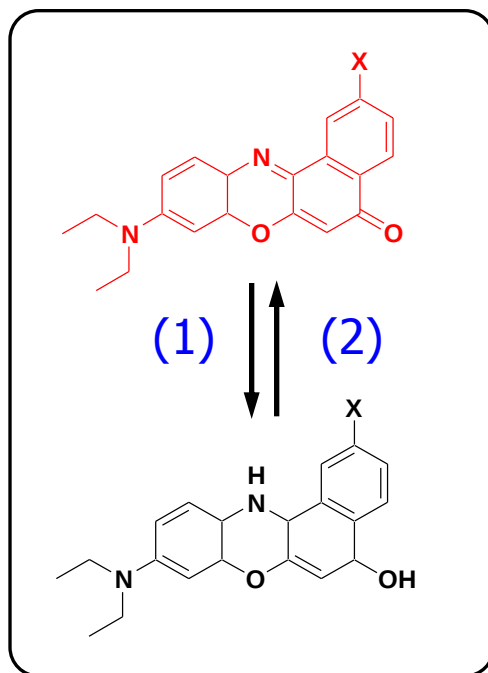
+



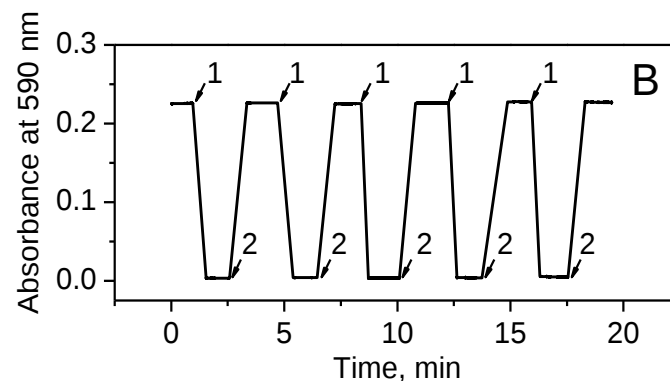
ON/OFF switching

state ON
Fluorescent
Red color

state OFF
non fluorescent
Colorless



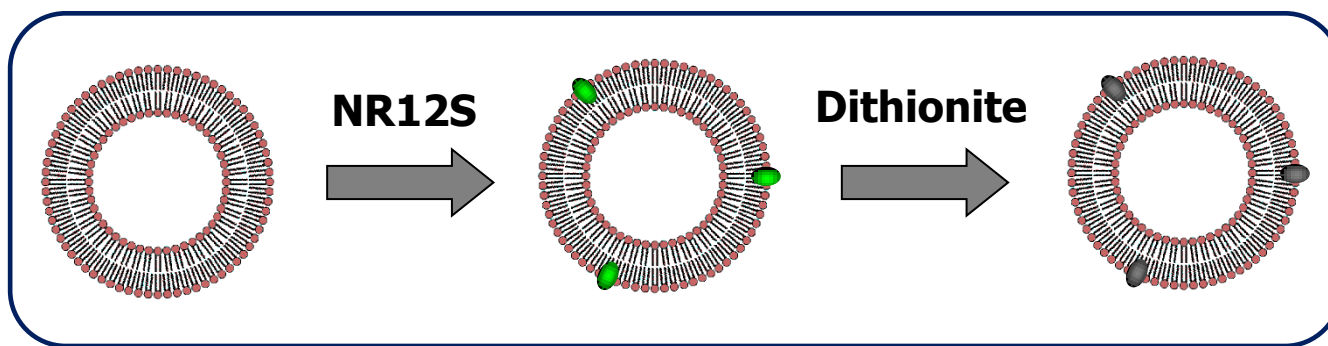
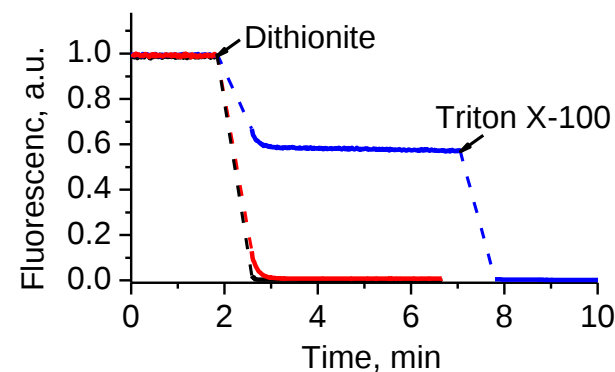
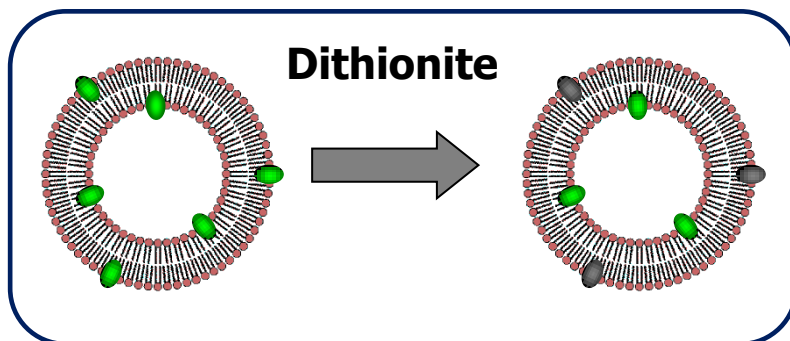
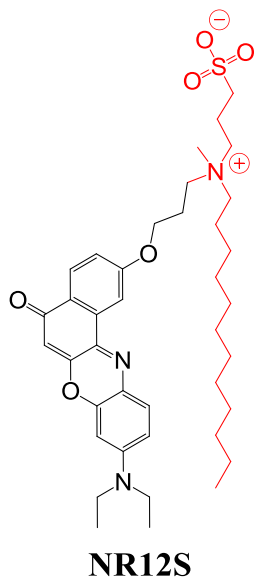
Switching in absorption of Nile Red



(1) addition of dithionite
(2) air bubbling

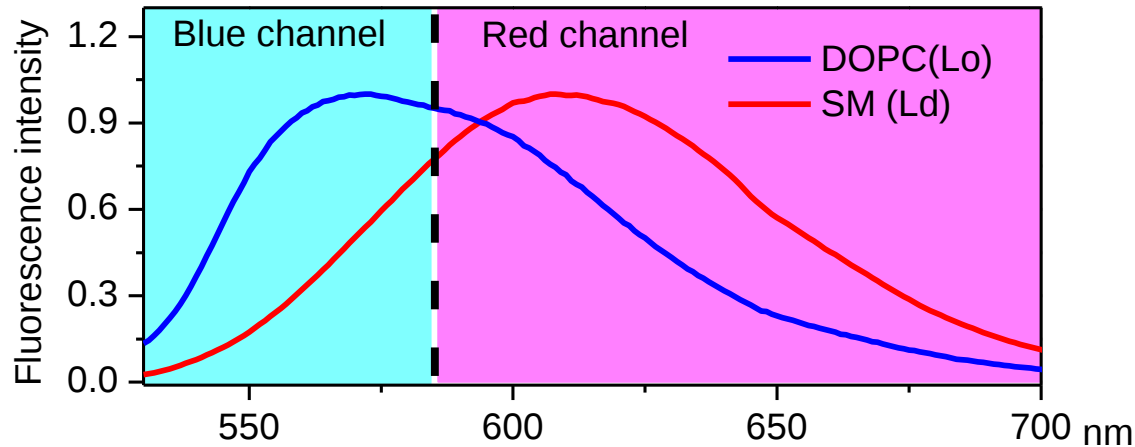
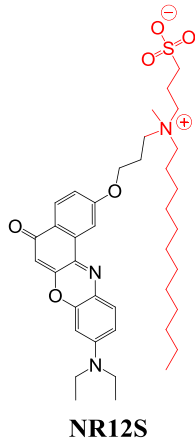
NR12S can be switched ON-OFF by dithionite and air

Selectivity to outer leaflet

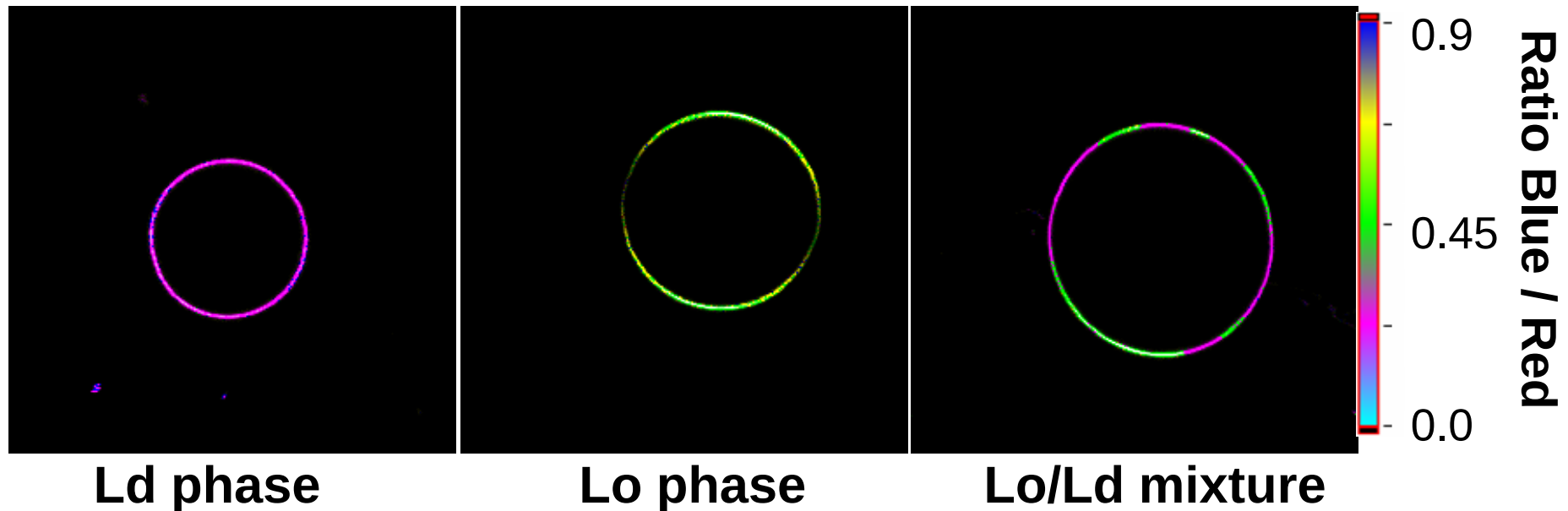
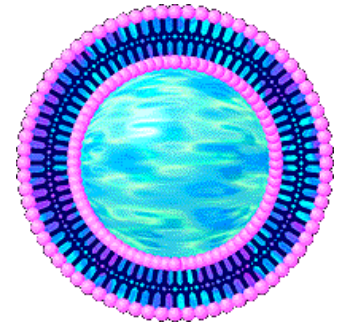


NR12S binds selectively outer membrane leaflet
without detectable flip-flop

Probing and imaging membrane phases in model membranes by NR12S

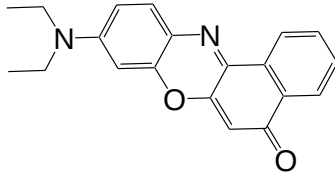


In liposomes

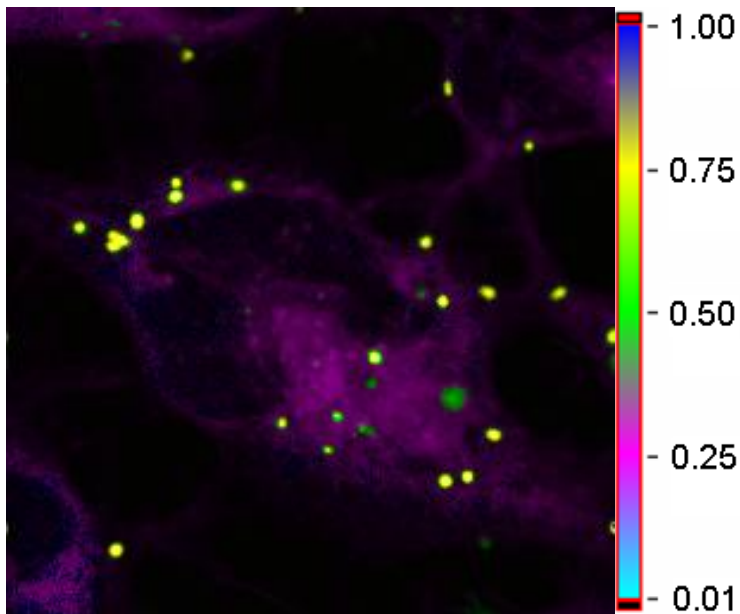
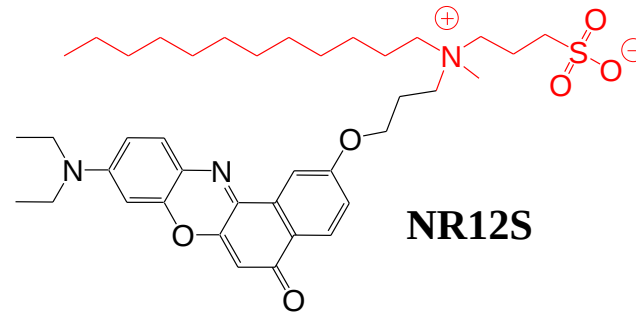


NR12S provides good color contrast of different phase domains

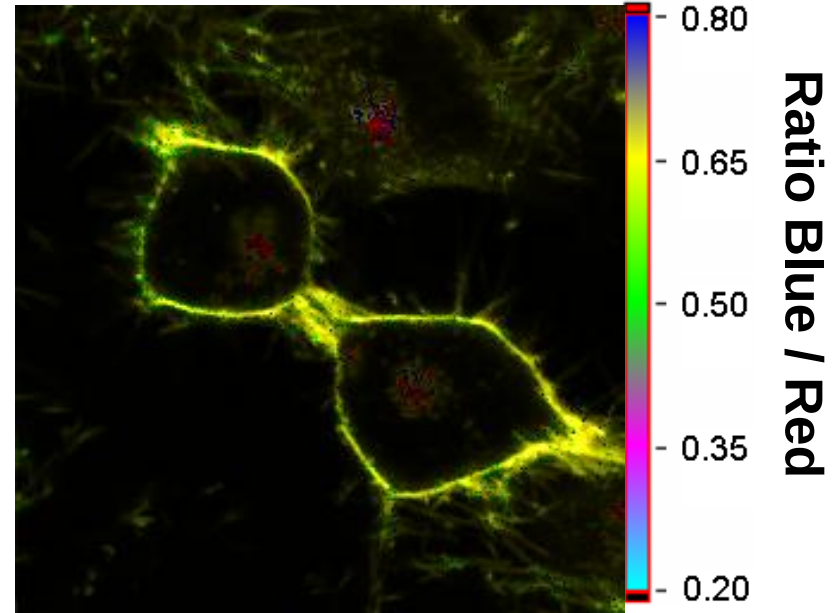
Selective staining of cell plasma membranes



Nile Red



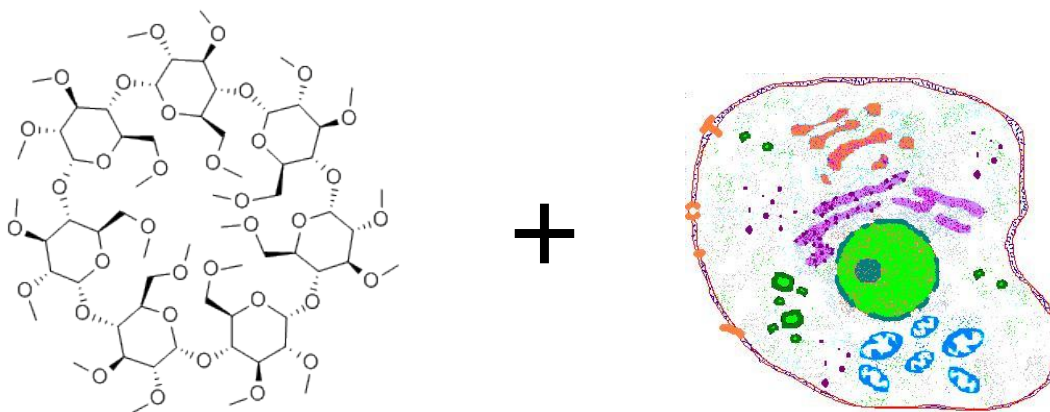
All cell + lipid droplets



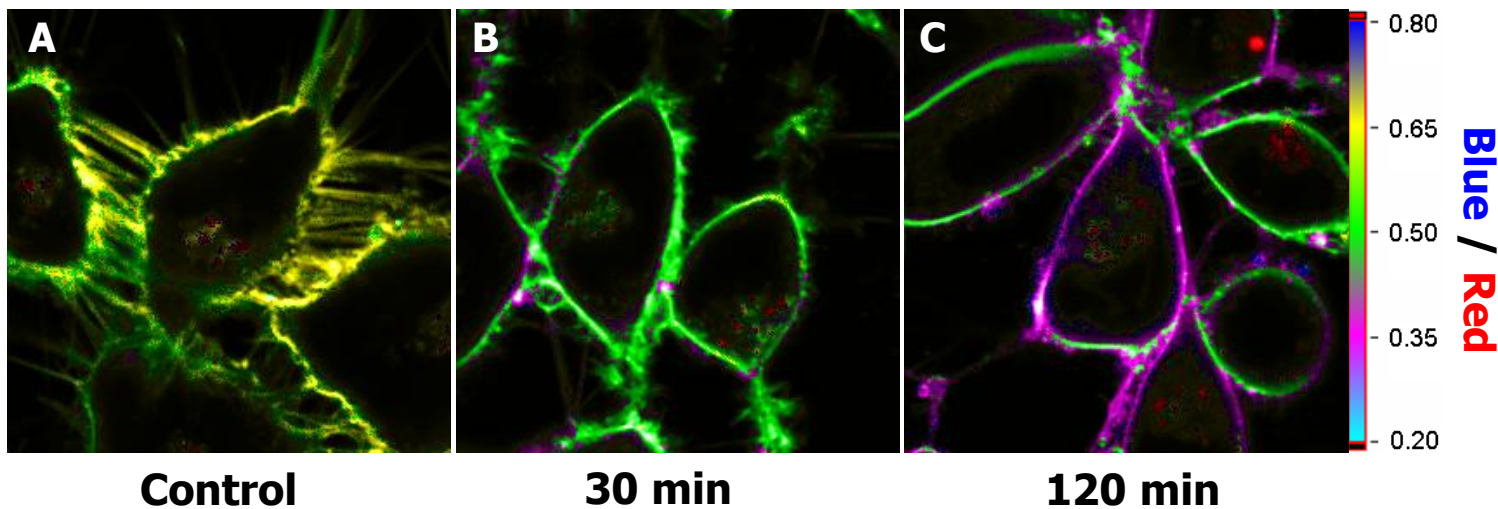
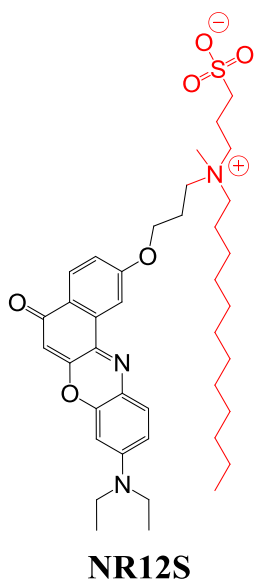
Cell membranes only!

NR12S binds exclusively cell plasma membranes

Color-Imaging of cholesterol content



Cholesterol extraction by Methyl- β -cyclodextrin

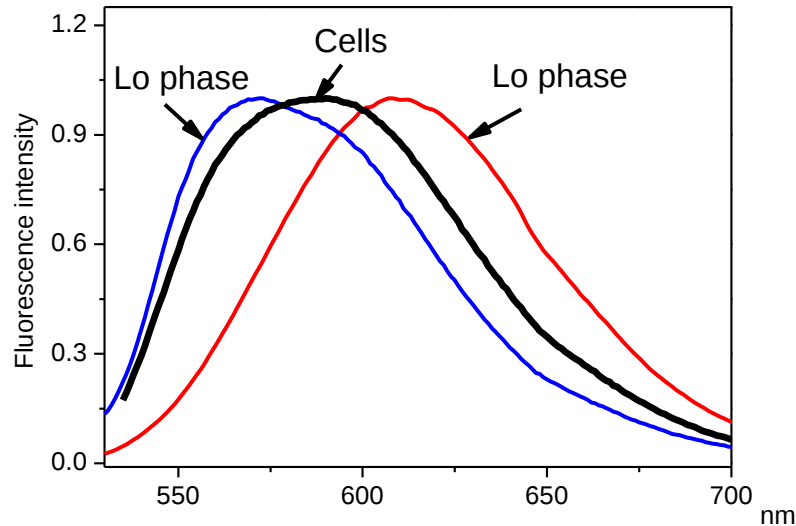


Kucharak et al., JACS 2010

NR12S is sensitive to cholesterol content in the membrane

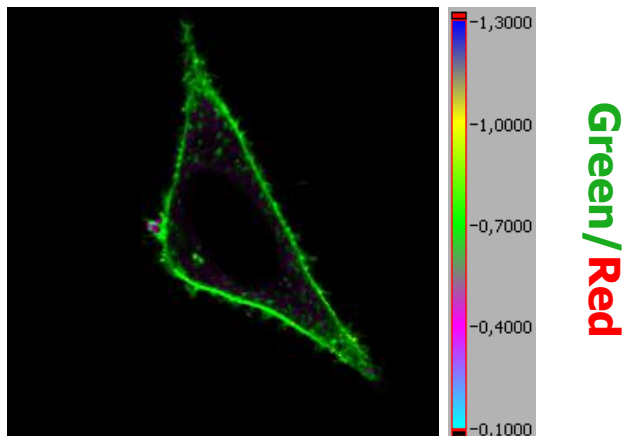
Where are the Rafts in Biomembranes?

LUVs and cells



- Intermediate spectrum Lo-Ld

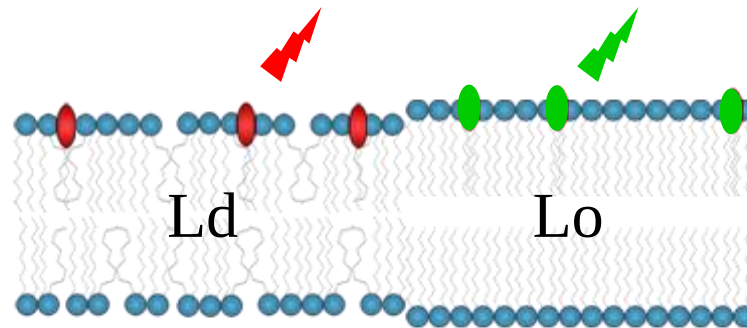
Intact cells



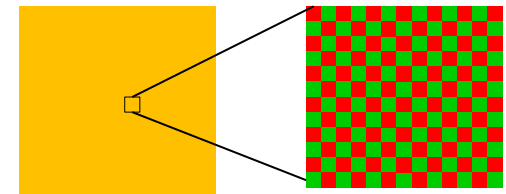
- But impossible to detect rafts on intact cells
- Limited temporal and spatial resolution of optical microscopy

Phase-specific Nile Red probe

- NR12S binds both phases

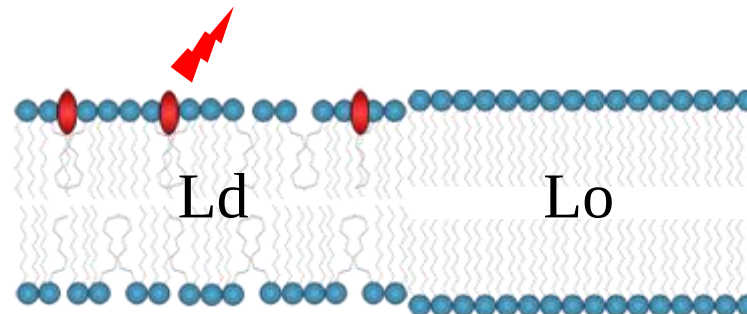


Colors mix up:

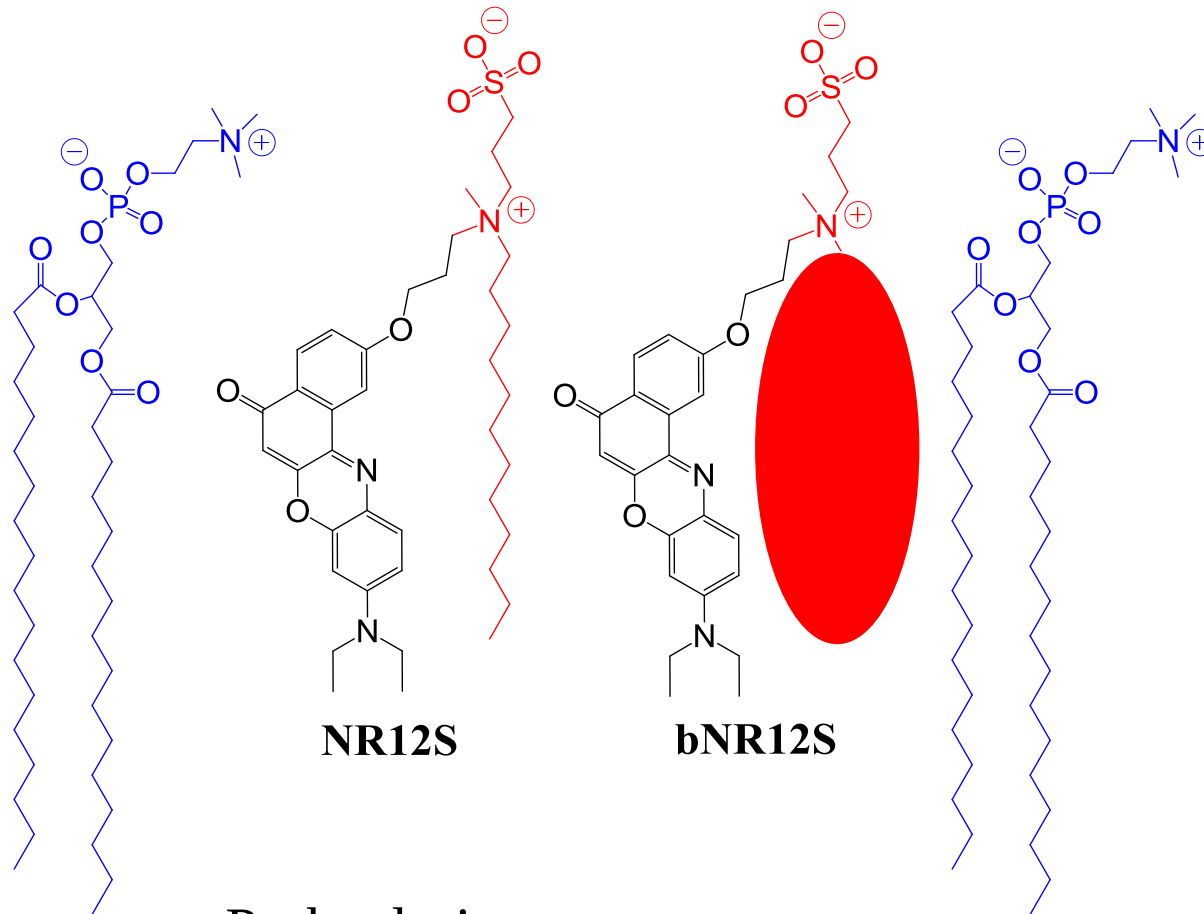


Zoom in

- “Bulky” NR12S analogue binding specifically to Ld phase



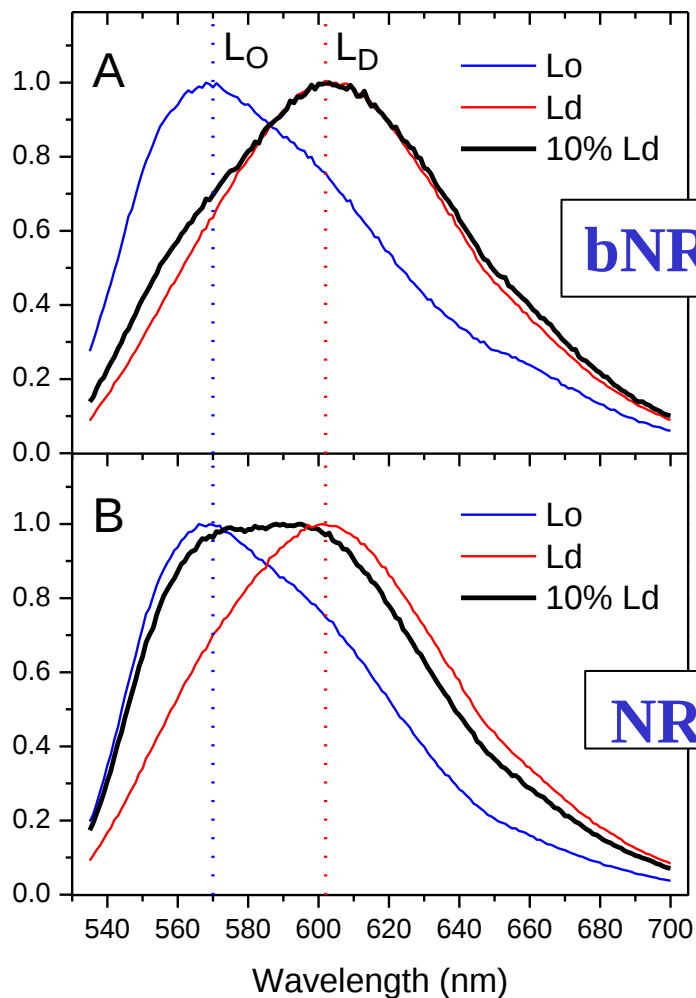
Design of the probes



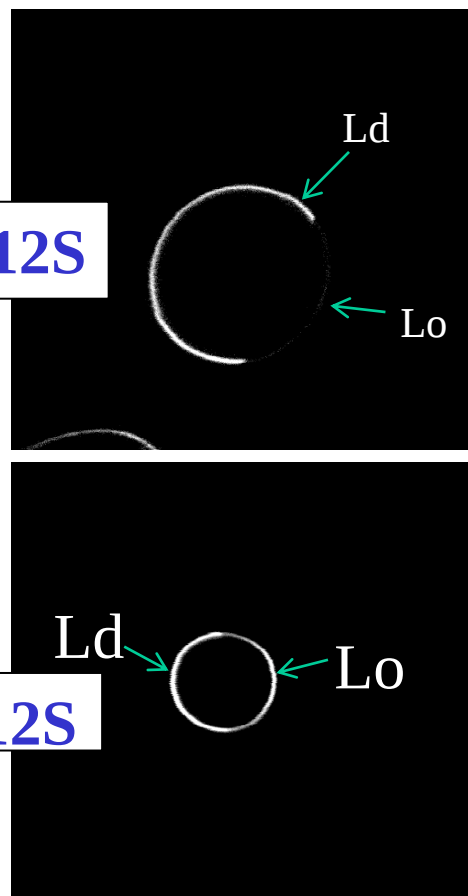
- Probe design:
 - Analogue of NR12S
 - A bulky alkyl chain

Validation in model membranes and cells

100-nm vesicles

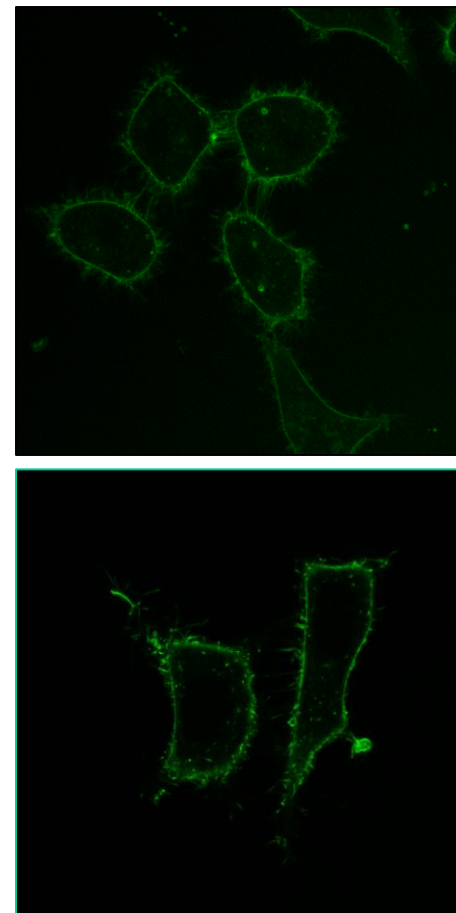


Giant vesicles



Lo – Sphingomyelin/Cholesterol,
Ld – DOPC/Cholesterol

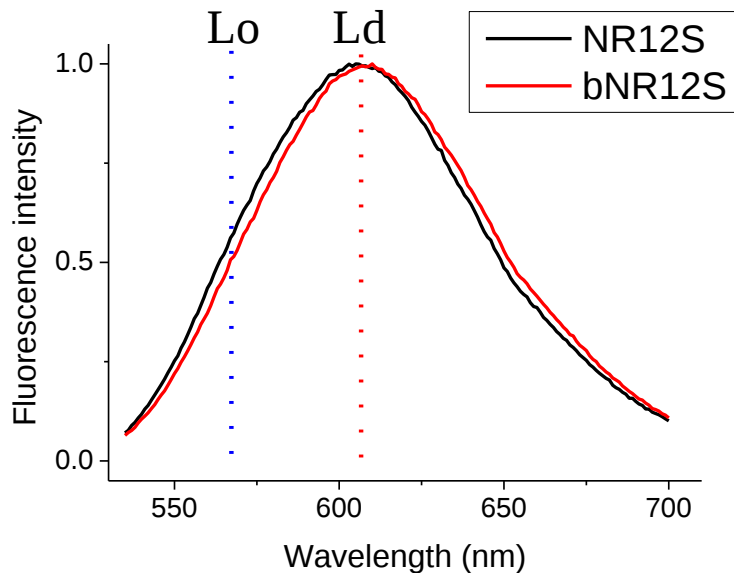
Cells



*HeLa cells stained with 0.4 μ M
of probes*

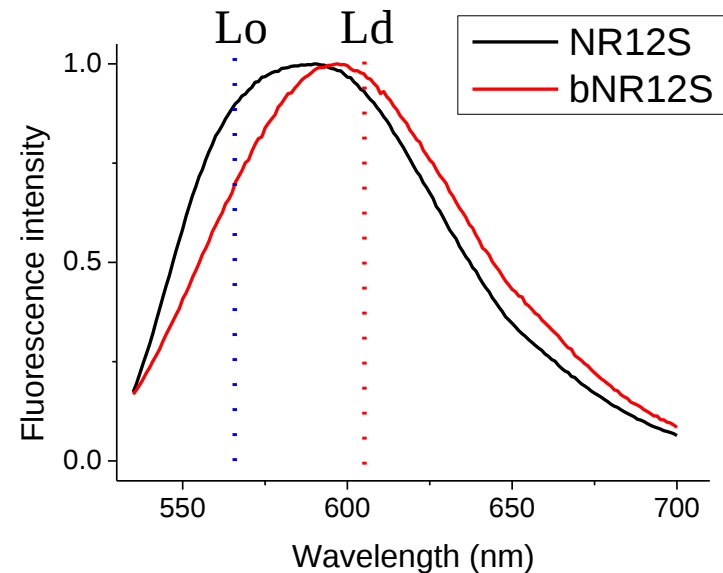
Comparison of NR12S and bNR12S in liposomes and HeLa cells

LUVs, Ld phase (DOPC)



Same spectra:
Homogeneous Ld phase

HeLa cells

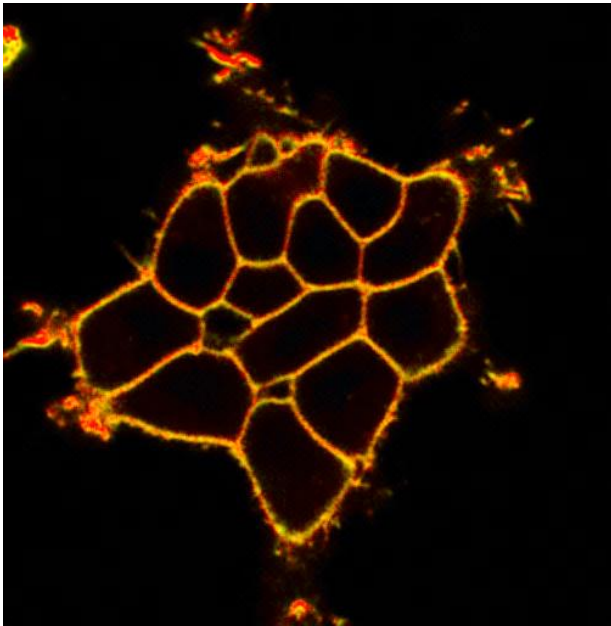


Different spectra:
bNR12S is excluded from Lo phase
NR12S partitions between Lo&Ld phases?
Coexistence of Ld&Lo phases!?

Direct observation of lipid rafts using phase-specific Nile Red probe

HEK293 cells stained with

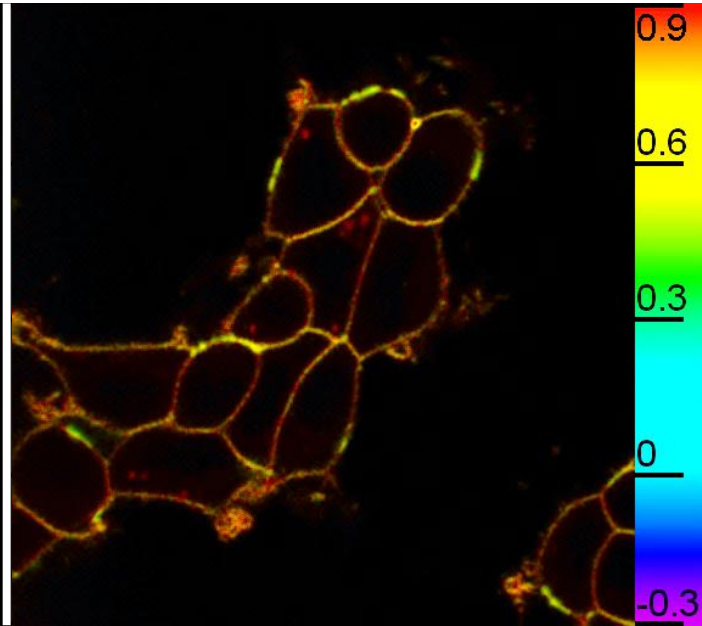
NR12S



Homogeneous: poor specificity to phases

bNR12S

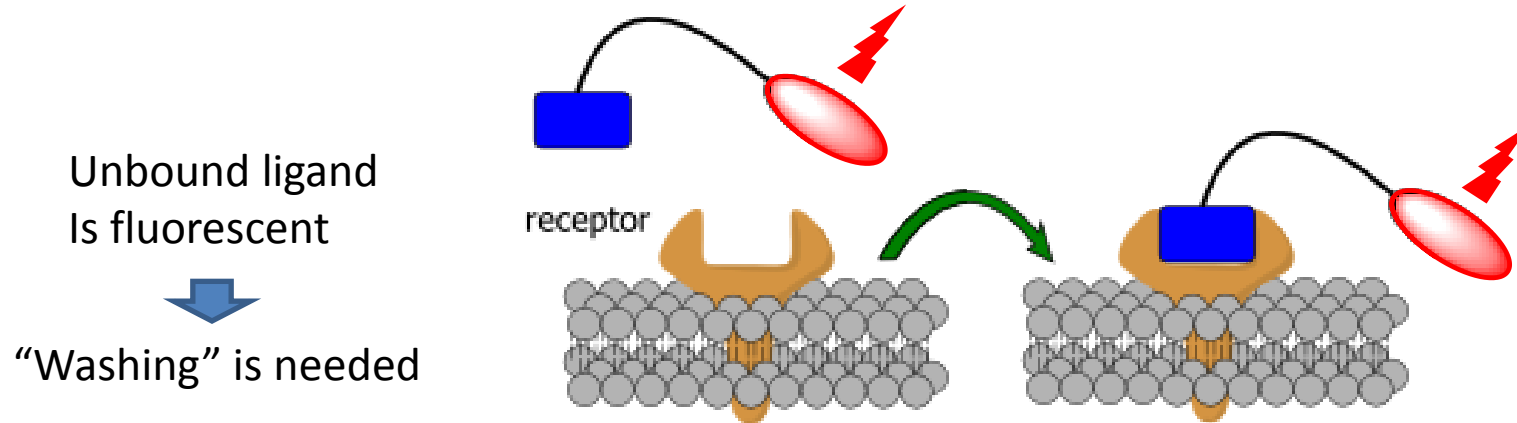
Green/Red



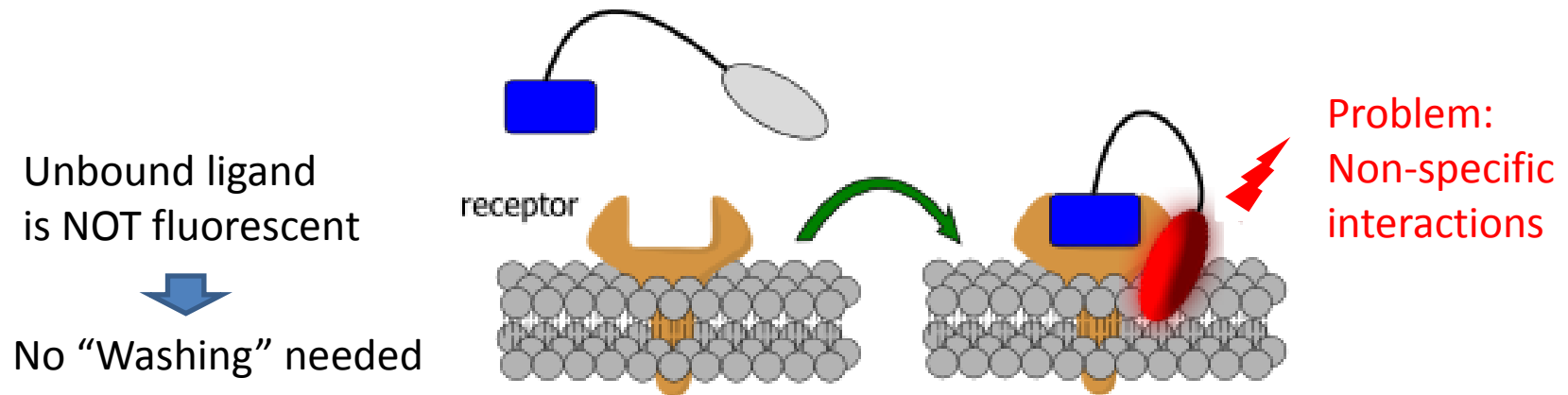
Domains of disordered phase can be distinguished by intensity and color!

Detection of membrane receptors

Classical Approach



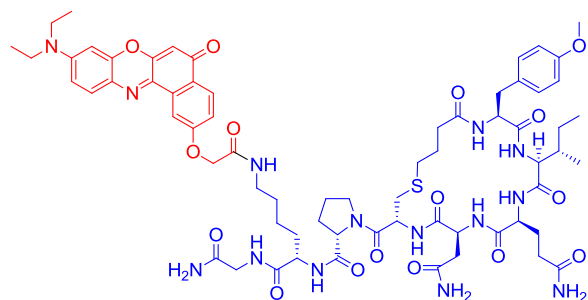
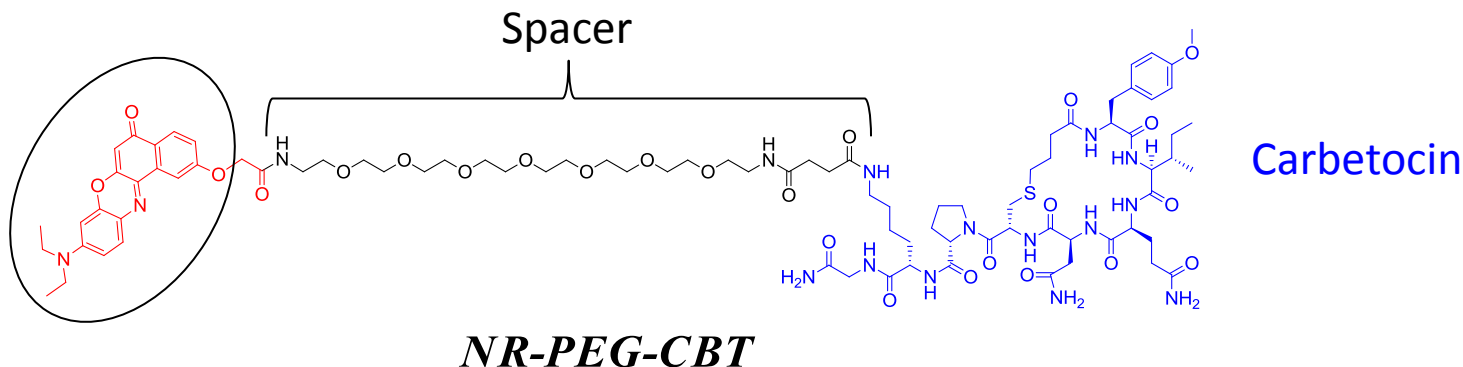
“Turn-ON” Approach



« Turn – on » ligands for G protein-coupled receptors (oxytocin)

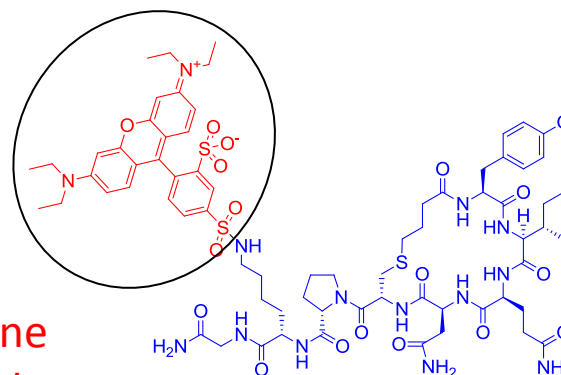
GPCRs is the target of ~ 40% of drugs on the market.

Nile Red
Turn-on dye:
Water – “OFF”
Oil – “ON”



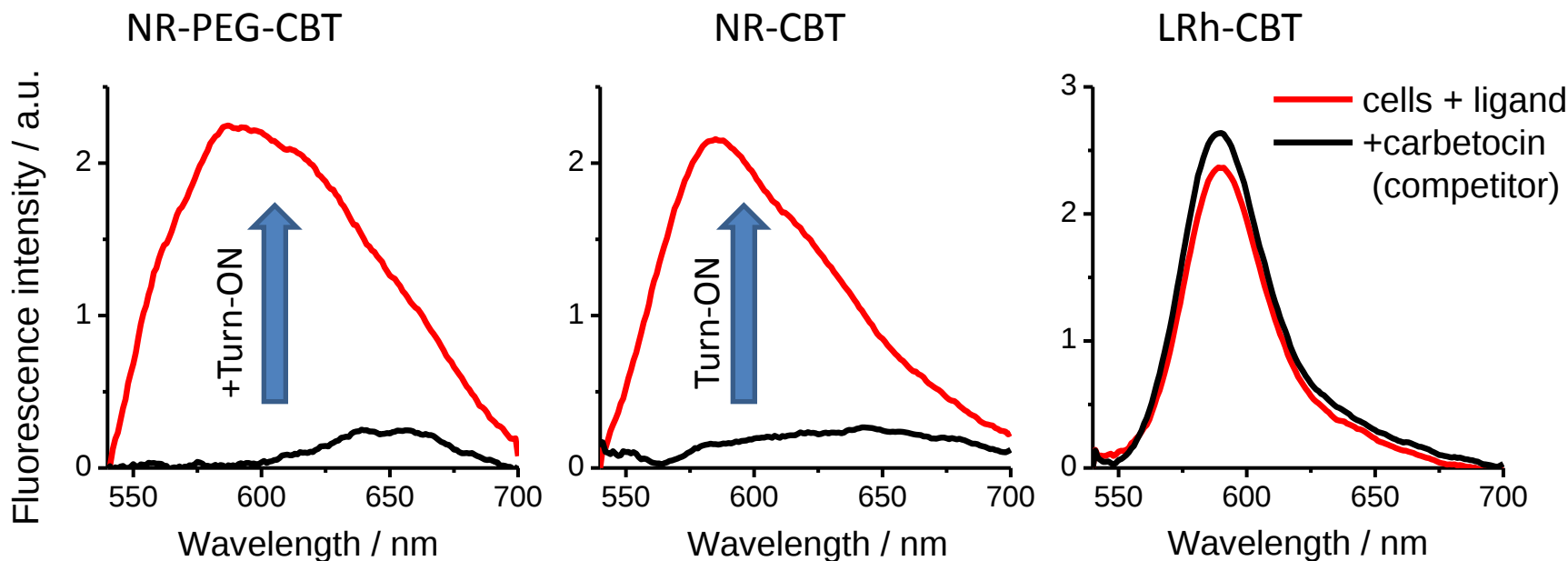
NR-CBT

Rhodamine
Classical dye
Always “ON”



LRh-CBT

Turn-on vs classical ligands in suspensions of HEK 293 cells



- ✓ New ligands turn on their fluorescence in the presence of receptor
- ✓ No effect is observed with classical ligand

Direct quantification: 29000 ± 5000 available receptors per cell
Independent radio-ligand method: 46000 ± 8000 receptors per cell

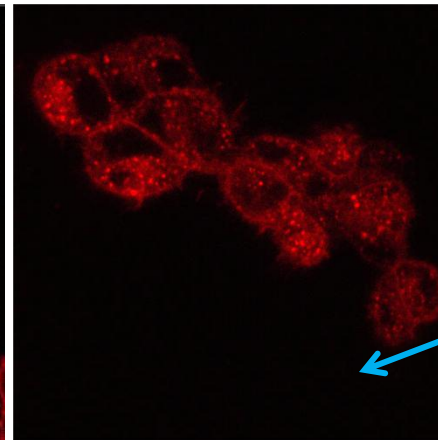
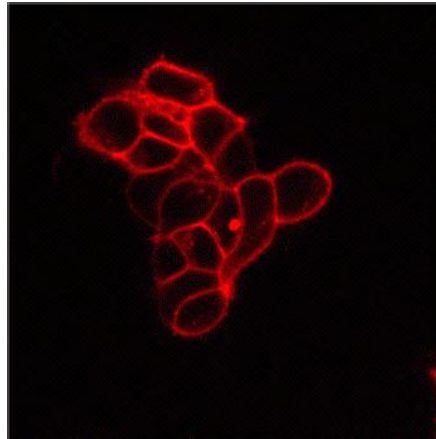
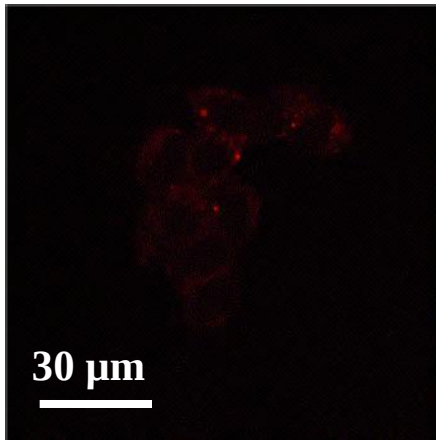
Turn-on vs classical ligand: Fluorescence microscopy

Cells without OTR
+20 nM ligand

Cells with OTR
+20 nM ligand

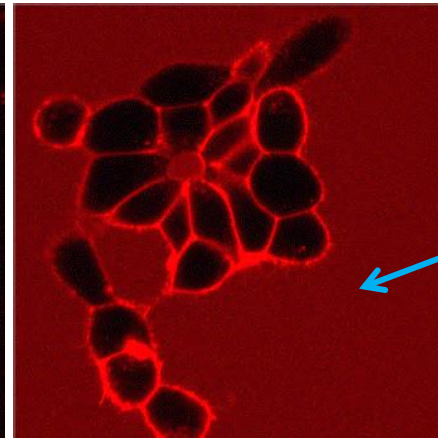
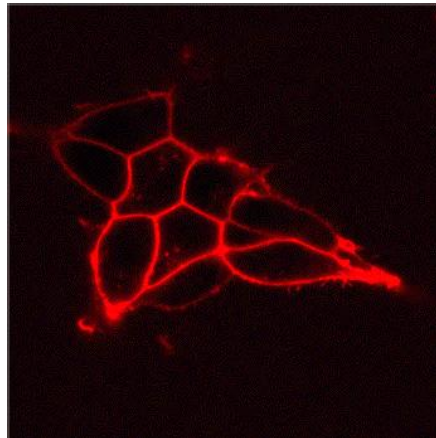
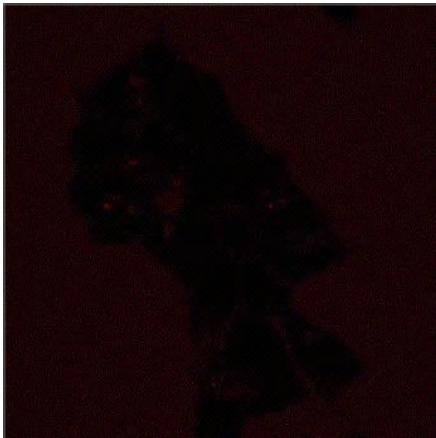
Cells with OTR
+100 nM ligand

Turn-ON
ligand



No
background

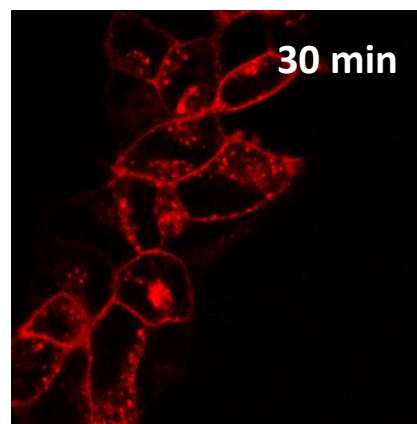
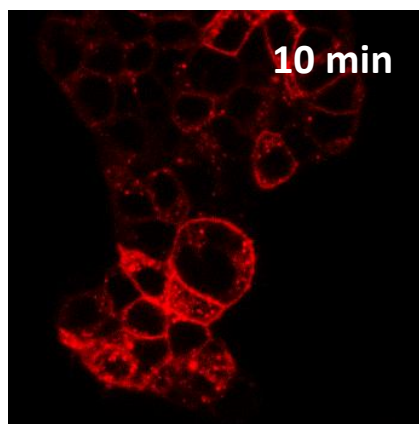
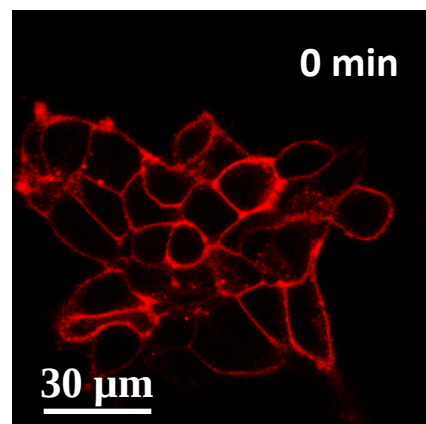
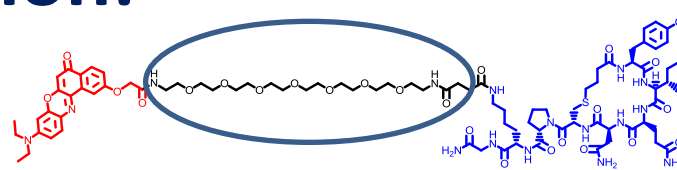
Classical
ligand



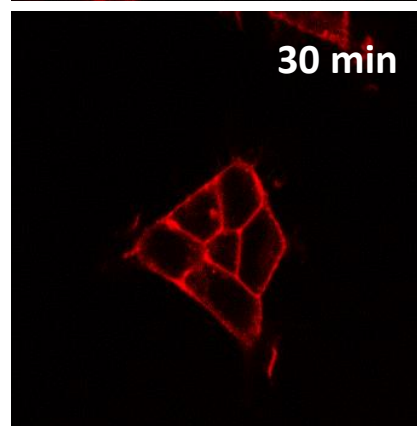
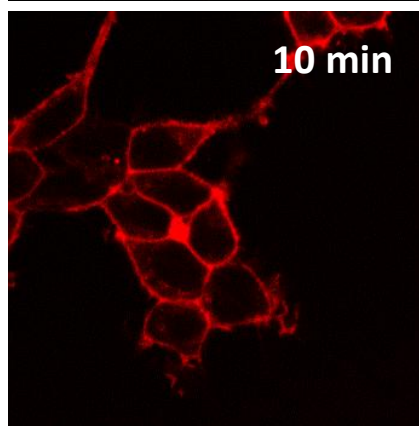
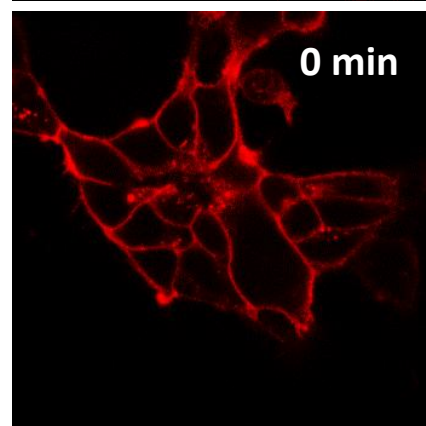
Strong
background

Turn-on ligand ensures receptor detection without background

Turn-on ligand internalization: Effect of spacer



Agonist
(with spacer)



Antagonist
(without spacer)

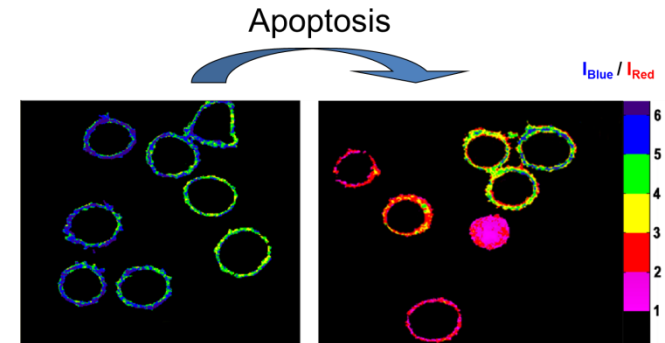
Long spacer preserves the agonist activity of carbetocin

CONCLUSIONS AND OUTLOOK

A tool kit for membrane research was developed based on environment-sensitive dyes:

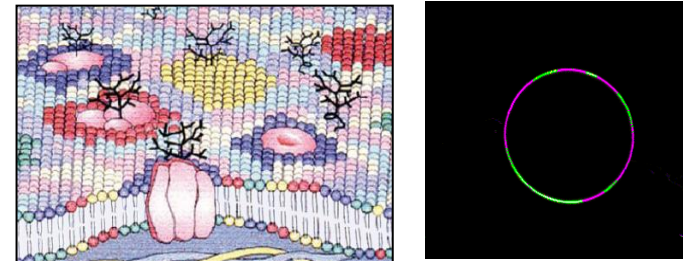
Probes for apoptosis

- ✓ Detection of transmembrane asymmetry
- ✓ Ratiometric response
- ✓ Suitable for screening pro-apoptotic agents
- ✓ In vivo applications?



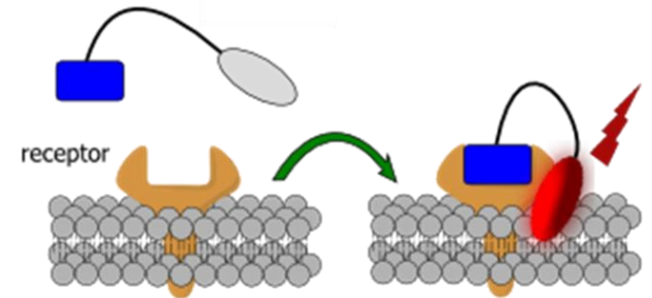
Probes for lipid order

- ✓ Imaging domains in model membranes
- ✓ Detection of cholesterol
- ✓ A phase-sensitive probe allows direct observation of lipid rafts in cells



Probes for membrane receptors

- ✓ Background-free detection of receptors
- ✓ Prospective for *in vivo* detection of overexpressed receptors in cancer cells



Acknowledgments

Lab members:

- Yves Mély
- Ludovic Richert
- Youri Arntz
- Pascal Didier
- Romain Vauchelles
- Rémy Kreder
- Zeinab Darwich
- Oleksandr Kucherak
- Sule Oncul



Collaborators:

- Julia Karpenko, Dominique Bonnet, Marcel Hibert, UMR 7200 (Strasbourg)
- Alain Wagner, Nicolas Anton, Thierry Vandamme, UMR 7199 (Strasbourg).
- Vasyl Pivovarenko and Alexander Demchenko (Ukraine)



ANR JC, ANR Blanc, Conectus,
ARCUS, Sanofi–Pasteur, etc.